

USE OF A CYCLOPENTENONE PROSTAGLANDIN FOR DELAYING FOR THE ONSET AND/OR PREVENTING THE CONTINUATION OF LABOUR

The present invention relates to agents for improving perinatal outcome in pre-term labour. In particular, the present invention relates to the use of prostaglandins to prevent and/or reduce an inflammatory response in the reproductive system of a female, thereby delaying the onset of labour.

Human pre-term labour, defined as spontaneous labour occurring prior to 37 weeks of gestation (with 39 weeks being term) continues to be a major problem, particularly in developed countries. Preterm birth occurs in 5-10% of all pregnancies but is associated with 70% of all neonatal deaths and up to 75% of neonatal morbidity (Rush *et al.*, 1976). Premature neonates are at high risk of cerebral palsy, developmental delay, visual and hearing impairment and chronic lung disease.

During pregnancy, the uterus is maintained in a state of non-contractile quiescence whilst the cervix remains firm and closed. With the onset of labour, the cervix needs to become softer and to offer low resistance to force applied and have fibres which move under tension. The uterus also needs to begin contracting.

Both at term and preterm, the biochemistry of labour resembles an inflammatory reaction and there is accumulating evidence pointing to a pivotal role for pro-inflammatory cytokines and prostaglandins (PGs) in the labour process. Interleukin-1 β (IL-1 β) levels are elevated in amniotic fluid (Romero *et al.*, 1990), gestational membranes (Keelan *et al.*, 1999; Elliot *et al.*, 2001) and the lower uterine segment (Maul *et al.*, 2002) at term labour, and may contribute to labour onset by stimulating IL-8 and PG synthesis (Mitchell *et al.*, 1990; Brown *et al.*, 1998). PGs increase in maternal urine and blood and in fetal membranes in association with labour (Sato *et al.*, 1979; Skinner and Challis, 1985). PGE₂ stimulates uterine contractions

(Dyal and Crankshaw, 1985), indirectly increases fundamentally dominant myometrial contractility by upregulation of oxytocin receptors and synchronisation of contractions (Garfield *et al.*, 1990), and acts in concert with IL-8 to remodel the cervix (reviewed in Kelly, 2002).

5

The onset of labour is associated with activation of the Nuclear Factor Kappa B (NF κ B) transcription factor system in the amnion which plays a role in the expression of pro-inflammatory genes such as interleukin-8 (IL-8), interleukin-6 (IL-6) and cyclo-oxygenases 1 and 2 (COX-1 and COX-2).

10 COX genes are also referred to as prostaglandin H synthase or PG synthase. The resulting inflammatory infiltrate (mediated by the cytokines) and increase in prostaglandin synthesis (mediated by the cyclo-oxygenases) leads to cervical ripening, fetal membrane rupture and myometrial contractions.

15

Five members of the NF- κ B/Rel family have been identified in mammals: NF- κ B1 (p50 and its precursor p105), NF- κ B2 (p52 and its precursor p100), p65 (RelA), c-rel, and Rel B. These proteins share a structurally conserved amino-terminal region termed the Rel homology domain (RHD). The RHD

20 is responsible for dimerisation, DNA binding, and interaction with the inhibitors of kappa B (I κ B) proteins. It also contains a nuclear localisation signal (NLS). In its active DNA-binding form NF- κ B consists of heterogeneous dimers of various combinations of NF- κ B subunits: each member of the NF- κ B family, except for Rel B, can form homodimers, as
25 well as heterodimers with one another. The p65, c-rel and Rel B proteins contain a carboxy-terminal non-homologous transactivation domain, which activates transcription from κ B sites in target genes; in contrast, p50 and p52 proteins lack a transactivation domain. The various NF- κ B dimers exhibit different binding affinities for specific κ B sites (Kunsch *et al.*, 1992,

Phelps *et al.*, 2000), and differentially stimulate transcription through distinct κ B elements (Lin *et al.*, 1995).

In resting cells, NF- κ B dimers are normally sequestered in an inactive form in the cytoplasm by association with the inhibitory I κ B proteins, which include I κ B α , I κ B β and I κ B ϵ . The I κ Bs are characterised by the presence of multiple ankyrin repeats which mediate binding to the RHD and mask the NLS of NF- κ B.

The major NF- κ B signalling pathway, which is activated by pro-inflammatory stimuli and LPS, targets I κ B α - and I κ B β -bound NF- κ B (for review see Li and Verma 2002). p50/p65 dimers are the most abundant form of NF- κ B in most cell types, and activation of I κ B α -bound p50/p65 dimers is the best characterised pathway. In this 'classical' pathway, diverse stimuli trigger signal transduction cascades that ultimately converge on the activation of a specific I κ B kinase (IKK). The IKK complex consists of several proteins, the main ones being IKK α (IKK1), IKK β (IKK2), and NF- κ B essential modulator (NEMO or IKK γ). The activated IKK complex phosphorylates I κ B α at serines 32 and 36, which results in the poly-ubiquitination of I κ B α at lysines 21 and 22. This modification targets I κ B α for rapid degradation by the 26S proteasome. The degradation of the I κ B inhibitor exposes the NLS of NF- κ B resulting in translocation of the p50/p65 dimer to the nucleus where it can bind to κ B sites in the promoter of target genes and promote transcription.

Most stimuli cause only the transient activation of NF- κ B. The critical inhibitory step in NF- κ B inactivation involves binding of newly synthesised I κ B α to NF- κ B in the nucleus. I κ B α is quickly resynthesised following its degradation. The newly synthesised I κ B α is localised in the nucleus and displaces NF κ B from its DNA binding sites. I κ B α contains leucine-rich

nuclear export sequences (NES) (Johnson *et al* 1999), which then enable it to transport NF- κ B back to the cytoplasm, thereby completing an autoregulatory post-induction repression.

- 5 In many cells nearly half of the NF- κ B is sequestered by the other major I κ B isoform, I κ B β (Whiteside *et al.*, 1997). In contrast to I κ B α , I κ B β is not NF- κ B inducible and does not exert a rapid post-induction repression of NF- κ B activity. Rather, I κ B β has been implicated in persistent NF- κ B activation. Prolonged exposure to certain stimuli, such as LPS, leads to the
- 10 long-term induction of NF- κ B activity despite high levels of newly synthesised I κ B α . Following stimulus-induced degradation, the newly synthesised I κ B β is un-phosphorylated and, in contrast to I κ B α or the constitutively phosphorylated I κ B β , can interact with NF- κ B bound to target promoters without displacing it from the DNA (Suyang *et al.*, 1996).
- 15 This interaction of un-phosphorylated I κ B β with DNA-bound NF- κ B is thought to protect NF- κ B from nuclear export, and thus inhibition, by I κ B α , and the outcome is a sustained NF- κ B response.

- PGs are a family of biologically active molecules having a diverse range of
- 20 actions depending on the prostaglandin type and cell target. There is considerable evidence to support a central role for PGs in human parturition. Labour is associated with increased PG synthesis within the uterus (Turnbull 1977) particularly from the fetal membranes (Skinner and Challis 1985). PGs act to mediate cervical ripening and to stimulate uterine
- 25 contractions (Cranckshaw and Dyal 1994) and indirectly to increase fundamentally dominant myometrial contractility by up-regulation of oxytocin receptors and synchronisation of contractions (Garfield *et al* 1990). PG synthesis in amnion, chorion-decidua and myometrium increases with labour (for a review, see Bennett and Slater 1996). Chorion
- 30 prostaglandin dehydrogenases are thought to protect the uterus from basal

prostaglandin synthesis during pregnancy but are down-regulated at term. Deficiency of prostaglandin dehydrogenase in chorion has been associated with pre-term labour (van Meir 1996, 1997).

- 5 Accordingly, inhibition of prostaglandin synthesis is an effective method of preventing or arresting pre-term labour (Keirse, 1995). Conversely, prostaglandins have been administered to induce labour as a means to terminate pregnancy (Ganstrom *et al.*, 1987).
- 10 Most PGs bind to prostanoid receptors localised on the cell surface and act through second messenger systems (Narumiya, 1995). However, PGD₂ metabolites are actively incorporated into the nuclei of cells (Narumiya *et al.*, 1987) and can exert their effects through direct interactions with nuclear receptors. Peroxisome proliferator-activated receptors (PPARs) are ligand-
- 15 activated transcription factors belonging to the nuclear receptor superfamily. They exist in three distinct forms, PPAR- α , PPAR- δ , and PPAR- γ , which form heterodimers with the retinoic X receptor (RXR) and bind to PPAR response elements (PPREs) in the promoter of target genes to induce transcription. PPAR- γ can also repress gene transcription by
- 20 negatively interfering with the NF- κ B, AP-1, STAT and C/EBP pathways (Zhou *et al.*, 1999; Subbaramaiah *et al.*, 2001; Takata *et al.*, 2002; Suzawa *et al.*, 2003).

- The aetiology of pre-term labour is multi-factorial but bacterial infection is
- 25 believed to play an important role, especially at earlier gestational ages (for review see Romero *et al.*, 2002). A growing body of epidemiological data suggests that intrauterine infection is an important cause of brain injury in infants born before 32 weeks of gestation. During ascending intrauterine infection, micro-organisms can stimulate the production of pro-
- 30 inflammatory cytokines, such as tumour necrosis factor α (TNF α) and IL-1 β , as well as PGs and other inflammatory mediators, resulting in the

premature onset of labour. Intrauterine infection/inflammation has also been identified as a key contributor to the development of cerebral palsy (CP) and schizophrenia (Urakubo *et al.*, 2001; Gibson *et al.*, 2003), and, although CP does occur in term infants, the risk of CP is strongly associated with prematurity (Dammann *et al.*, 1999).

In addition, inflammatory responses caused by mechanical stretching of the uterus may contribute to the onset of labour. Mechanical stretching of the uterus occurs to an extent as a normal part of pregnancy and may be responsible for some of the biochemical changes which occur near to term and which cause the normal onset of labour at term. In the context of preterm labour, mechanical stretch may occur where the uterus is overdistended by multiple pregnancy or by excess amniotic fluid (clinically termed hydramnios or polyhydramnios). There may also be more local stretch of the lower segment of the uterus, the cervix and overlying fetal membranes in cases where there is cervical weakness (clinically termed cervical incompetence). Stretch leads to an increase in the production of a series of 'labour-associated' proteins including COX-2 (which then increases prostaglandin synthesis), cytokines such as IL-8 and IL-1b and the oxytocin receptor. Increased prostaglandin and cytokine productions causes cervical ripening or further cervical ripening (and may lead to neonatal brain injury). Prostaglandins and OTR receptor lead to uterine contractions.

Obstetric management of pre-term labour is still largely reactive and centred on the use of drugs intended to inhibit contractions to delay delivery. This was thought to be principally dependent upon gestational age leading to the concept that prolongation of the pregnancy will always improve outcome. However, there is now growing evidence that the mechanisms leading to pre-term birth also cause fetal cerebral damage. Characteristically, damage is localised to the white matter, involving both a diffuse astrogliosis with subsequent loss of myelin-producing oligodendrocytes, as well as

multifocal necroses resulting in cystic change (periventricular leucomalacia, PVL). Such lesions lead to cerebral palsy in 60–90% of affected infants (described in Vlope, 2001).

5 There are currently no drugs available which will safely and effectively inhibit pre-term contractions. The most commonly used agents, β -sympathomimetics such as Ritodrine, Salbutamol and Terbutaline, cause significant maternal cardiovascular, respiratory and metabolic side effects and may lead to pulmonary oedema, cardiac failure and maternal death.
10 Furthermore they are subject to tachyphylaxis and become ineffective after 24 to 48 hours. Meta-analysis of randomised controlled trials has shown that the value of β -sympathomimetics is only in the temporary delay of labour to allow *in utero* transfer or administration of steroid to improve fetal lung surfactant production.

15 Other than the antenatal administration of corticosteroids, no obstetric interventions affect neonatal outcome although improvements in neonatal intensive care have dramatically increased survival rates. Commonly used agents are dexamethasone or betamethasone. Antenatal administration of
20 corticosteroids improves the outcome for the pre-term neonate since it reduces the incidence and severity of respiratory distress syndrome, intracranial haemorrhage and necrotising enterocolitis. One function of corticosteroids is to mature the fetal lung, which leads to an increase in surfactant production and therefore prevents or reduces the severity of
25 neonatal respiratory problems. Such agents are known to those skilled in the art.

Current obstetric management of pre-term labour (or threatened pre-term labour or pre-term premature rupture of membranes) is to attempt to delay
30 delivery using ‘tocolytic’ drugs to allow time for steroid administration.

Typically, effective tocolytic drugs are oxytocin receptor antagonists, calcium channel blockers, sympathomimetics and nitric oxide donors.

A commonly used oxytocin receptor antagonist is Atosiban, that functions
5 by blocking the oxytocin receptor, thereby preventing activation of the
receptor by endogenous oxytocin that stimulates uterine contractions. A
commonly used calcium channel blocker is Nifedipine, that functions to
block the influx of calcium into the myometrial cells, which is a
requirement for contractions to take place. A commonly used
10 sympathomimetic is Ritodrine, that functions by activating adrenergic
receptors on the myocyte cell membrane leading to phosphorylation and
down-regulation of the activity of myosin light chain kinase, an enzyme
essential for contractions. A commonly used nitric oxide donor is glyceryl
trinitrate, that functions by increasing myocyte cGMP thereby down-regulating
15 the activity of myosin light chain kinase, an enzyme essential for
contractions.

Indomethacin, a cyclo-oxygenase inhibitor, is effective in preventing the
contractions of pre-term labour. It is more effective in short term
20 prolongation of pregnancy than the β -sympathomimetics and, unlike
 β -sympathomimetics, it can reduce the risk of delivery pre-term (Keirse
1995). The use of indomethacin is limited by fetal side effects.
Indomethacin reduces fetal urine output and constriction of the ductus
arteriosus (Moise *et al* 1995). Clinically significant ductal constriction
25 occurs only in a proportion, increasing with gestational age from 10% at 26
weeks to 50% at 32 weeks. Accordingly the use of indomethacin is limited
in clinical practice to use \leq 32 weeks, and to short courses (\leq 48 hours) after
which any effects on the constriction of the ductus have been shown to be
reversible (Tulzer *et al* 1991; Moise *et al* 1993; Respondek *et al* 1995).

Because of these side effects some obstetricians now use Sulindac, which appears to be equally good as a tocolytic (Carlon *et al* 1992) in place of indomethacin. Sulindac produces a smaller reduction in fetal urine output and minimal effect on ductal patency (Carlon *et al* 1992; Rasanen and
5 Jouppila 1995). However, Sulindac is far from an ideal choice of tocolytic agent.

Accordingly, new agents or regimens capable of reducing and/or preventing an inflammatory response in the reproductive system of a female are highly
10 desired. Such medicaments or approaches would allow the treatment of pathogenic infection within the reproductive system of a female and/or delay pre-term delivery without causing injury to the fetus/neonate.

In light of the above, the present inventors have surprisingly discovered that
15 prostaglandins can be used to delay the onset and/or prevent the continuation of labour in a female.

Thus, in a first aspect, the present invention provides the use of a cyclopentenone prostaglandin in the manufacture of a medicament for
20 delaying the onset and/or preventing the continuation of labour in a female.

Preferably, this is achieved by preventing and/or reducing an inflammatory response in the reproductive system of a female.

25 The invention stems from the unexpected finding that the cyclopentenone prostaglandins, such as 15-deoxy- $\Delta^{12,14}$ prostaglandin J₂ (15-dPGJ₂) and prostaglandin A₁ (PGA₁), inhibit and/or reduce NF κ B activity within uterine cells of the female reproductive system. Thus, cyclopentenone prostaglandins provide a means for the inhibition and/or reduction of NF κ B
30 activity in the reproductive system of a female. Medicaments of the invention are believed to inhibit cytokine synthesis and inhibit the

biochemical processes of labour, thereby safely prolonging pregnancy. Accordingly, the present invention will improve obstetric management of pre-term labour as the onset of labour may be delayed without injuring the fetus/neonate.

5

The cyclopentenone prostaglandins are naturally-occurring substances that contain a cyclopentenone ring structure. The cyclopentenone ring is characterised by the presence of a chemically-reactive α,β -unsaturated carbonyl and is formed by dehydration of the cyclopentane ring of a precursor prostaglandin.

10

Generally, the first step in the biosynthesis of prostaglandins involves the intracellular release of arachidonic acid from plasma membrane phospholipids via the action of phospholipase A_2 . Arachidonic acid is then converted sequentially to PGG_2 and PGH_2 by the cyclo-oxygenase and peroxidase activities of the PGH synthases, PGH 1 and 2. The prostaglandins PGE_2 , PGD_2 and $PGF_{2\alpha}$ are subsequently synthesised from PGH_2 via the action of the PGE_2 , PGD_2 and $PGF_{2\alpha}$ synthase, respectively. The cyclopentenone prostaglandins, prostaglandin A_2 (PGA_2), prostaglandin A_1 (PGA_1) and prostaglandin J_2 (PGJ_2) are formed by dehydration of prostaglandin E_2 (PGE_2), prostaglandin E_1 (PGE_1) and prostaglandin D_2 (PGD_2), respectively. PGJ_2 is metabolised further to Δ^{12} -prostaglandin J_2 (Δ^{12} - PGJ_2), and 15-deoxy- $\Delta^{12,14}$ -prostaglandin J_2 (15-d PGJ_2).

20

Other unnatural or synthetic prostaglandins can be made by chemical synthesis. Total synthesis of prostaglandins was first accomplished by Corey in the 1960s (reviewed in Corey, 1991), and subsequently simplified by Suzuki *et al.* (1990). This latter scheme uses a C8 organometallic reagent for one side chain and a C7 acetylenic halide for the other side chain which are added to the desired chemical head-group. This synthesis is versatile and allows the synthesis of a variety of natural and unnatural

30

prostaglandins including the cyclopentenone prostaglandins. A general pathway for natural and chemical synthesis of prostaglandins and cyclopentenone prostaglandins is described in Straus and Glass (2001), the disclosure of which is incorporated herein.

5

Chemical modification of cyclopentenone prostaglandins using techniques known in the art of chemistry may alter the clinical effectiveness of the molecule. Such alterations may, for example, increase or decrease the stability or another characteristic of the cyclopentenone prostaglandin, to
10 give a desired change in activity. For example, modification of the 15C residue of cyclopentenone prostaglandins will reduce the metabolism of the compound, thereby increasing its half-life *in vivo*. Such modifications will be appreciated by those skilled in the art.

15 Thus, by “cyclopentenone prostaglandin”, we include any natural, unnatural or chemically-modified prostaglandin which has a cyclopentenone ring. Cyclopentenone prostaglandin is often abbreviated to “cyPG”. Especially preferred cyclopentenone prostaglandins include prostaglandin D₂ (PGD₂) and its metabolite 15-deoxy- $\Delta^{12,14}$ prostaglandin J₂ (15-dPGJ₂). Also
20 preferred is prostaglandin A₁ (PGA₁).

15-dPGJ₂ may be obtained from Cayman Chemical, 1180 East Ellsworth Road, Ann Harbour, MI 48108 USA (catalogue number 18570); 9,10-dihydro-15-deoxy- $\Delta^{12,14}$ -Prostaglandin J₂ may be obtained from Alexis
25 Biochemicals Ltd, PO Box 6757, Bingham, Nottingham, NG13 8LS, UK (catalogue number CAY-18590-M001). PGA₁ may be obtained from Alexis Biochemicals Ltd (address as above; catalogue number 340-045-M005).

30 By “onset of labour” and/or “continuation of labour” we include the biochemical and/or physiological changes associated with preparation of the

tissues of the female reproductive system for delivery. For example, the uterus increases in contractility and undergoes contractions. The cervix also ripens in readiness for delivery. Such changes are well known in the arts of obstetrics, gynaecology and midwifery and, for example, the Bishop's score
5 indicates the degree of cervical ripening (described in Herman *et al.*, 1993). By “delaying the onset of labour in a female and/or preventing the continuation of labour in a female” we include the meaning that at least one of these biochemical and/or physiological changes are delayed or prevented.

10 By “female” we include any female mammal such as human, or a domesticated mammal, preferably of agricultural significance including a horse, pig, cow, sheep, dog and cat. It is preferred if the female is a human female.

15 In a second aspect, the present invention provides the use of a cyclopentenone prostaglandin in the manufacture of a medicament for preventing and/or reducing an inflammatory response in the reproductive system of a female. Such medicaments are able to inhibit and/or reduce NFκB activity in uterine cells.

20

By “NFκB” we include homo- and heterodimers of RelA (p65), RelB, NFκB1 (p50), NFκB2 (p52) and cRel. The RelA (p65), RelB, NFκB1 (p50), NFκB2 (p52), and cRel genes and the sequence of the polypeptide products are described in Li *et al.* (2002).

25

By “NFκB activity” we include the activities of NFκB associated with the expression of genes controlled by any homo- or heterodimer of RelA (p65), RelB, NFκB1 (p50), NFκB2 (p52) or cRel of the NFκB transcription factor family. In particular, we include: nuclear translocation of NFκB which can
30 be measured, for example, by Western blotting analysis of nuclear and cytosolic cellular fractions for the protein of interest (described in

Sambrook *et al.*, 1989; Lee *et al.*, 2003); binding of NF κ B to target nucleic acid sequences (such as specific regions and sequences of DNA), which can be measured, for example, by Electro-Mobility Shift Assay (EMSA, as described in Dignam *et al.*, 1983; Lee *et al.*, 2003); and NF κ B-mediated
5 expression of target genes which can be measured, for example, by northern blotting and/or Western blotting (Sambrook *et al.*, 1989; Lee *et al.*, 2003). Methods for measuring these activities of NF κ B are well known by those skilled in the art of biochemistry and molecular biology.

- 10 By “uterine cells” we include any cells within the uterus of a female, or cells derived from the uterus of a female, particularly placental cells, amnion cells, myocytes, uterine and cervical fibroblasts, and maintained as a primary or transformed cell culture or line. These cell types are typically referred to as “gestational tissues”.

15

- Cultures of amnion cells may be prepared from tissue by separating the entire amnion, except for the part overlying the placenta, from the chorion, followed by separating amnion epithelial cells from fibroblasts and maintaining the epithelial cells using mammalian cell culture techniques
20 (Lee *et al.*, 2003). Myometrial cell culture may be prepared from tissue from the lower uterine segment, separating cells by incubation with Dispase and collagenase/elastase/DNAase solution and maintaining the myometrial cells using mammalian cell culture techniques (Pieber *et al.*, 2001). Techniques for the generation and maintenance of primary and transformed
25 mammalian cell cultures will be well known to those skilled in the relevant art.

- By “reproductive system of a female”, we include any cells and/or tissues and/or organs of a female directly or indirectly involved in the formation,
30 nourishment, maintenance and development of a neonate, embryo or fetus at any gestational stage during pregnancy. In particular we include the cells

and/or tissues of the uterus, placenta, amnion, chorion, decidua, cervix and vagina.

Preferably, the medicament is for preventing and/or reducing an
5 inflammatory response in the reproductive system of a female that is pregnant.

By “inflammatory response” we include biochemical and physiological changes associated with inflammation mediated by cells of the host’s
10 immune system. Such changes are known in the arts of human and veterinary medicine, immunology, molecular biology and biological science.

If a patient is detected clinically at being at high risk of preterm delivery,
15 because of detection of fibronectin in the vagina, identification of cervical shortening on ultrasound, the identification on clinical examination of cervical dilatation, or the onset of contractions then there is a high risk that there may be inflammation within the uterus. Other clinical measures of inflammation within the uterus are maternal temperature, white blood cell
20 count, serum c-reactive protein concentrations and amniotic cytokine concentrations (taken at amniocentesis) which suggest a high risk of inflammation within the uterus if abnormal. Methods for measuring such changes will be well known to those skilled in the art.

25 By “pregnant”, we include the meaning that the female is carrying a fertilised egg in the uterus, or an embryo or neonate or fetus at any stage of gestational development.

Preferably, the present invention provides a use wherein the female is
30 human and the duration of pregnancy is more than approximately 13 weeks

of human pregnancy. More preferably, the duration of pregnancy is approximately between 20 and 32 weeks.

5 Preferably, the medicament reduces and/or prevents an inflammatory response in the reproductive system of a female associated with the onset or continuation of labour. The biochemical and physiological changes associated with the onset or continuation of labour have been mentioned above.

10 There are many situations where it is useful to substantially prevent or reduce at least one of the changes in the female reproductive system associated with the onset or continuation of labour. For example, it is well known that certain groups of pregnant females are at high risk of pre-term labour. Females that have had one or more instances of pre-term labour
15 previously are at considerably higher risk of a further pre-term labour when pregnant. An increased risk of pre-term labour can also be determined by measuring oncofetal fibronectin levels and by cervical examination using methods well known in the art.

20 It is also useful to prevent or reduce at least one of the changes in the female reproductive system associated with the continuation of labour, particularly uterine contractions, temporarily in circumstances where this is desirable. For example, it may be desirable temporarily to inhibit uterine contractions during labour in order to clear the fetal lungs or in order to transfer the
25 female from one place to another. It is often desirable to transfer the female to a more suitable place where better care is available for her and the offspring.

It is also useful to substantially prevent for a considerable duration pre-term
30 labour using the method of the invention. In particular, it is useful to inhibit

pre-term uterine contractions from the time when they first occur (or soon thereafter) until the normal time of delivery.

Preferably, the medicament reduces and/or prevents an inflammatory
5 response in the reproductive system of a female associated with infection by a pathogenic agent

More preferably, the pathogenic agent is viral, bacterial or fungal.

10 Preferably, the medicament reduces and/or prevents an inflammatory response in the reproductive system of a female associated with stretch of the uterus.

By “stretch of the uterus” we include mechanical stretching of the uterus
15 occurring where the uterus is overdistended by multiple pregnancy or by excess amniotic fluid (clinically termed hydramnios or polyhydramnios). There may also be more local stretch of the lower segment of the uterus, the cervix and overlying fetal membranes in cases where there is cervical weakness (clinically termed cervical incompetence).

20

Preferably, the medicament reduces and/or prevents one or more of the following conditions: pre-term labour; pathogenic infection; cervical ripening, uterine contractions.

25 By “pre-term labour”, we include the meaning of spontaneous labour occurring before the usual calculated time for delivery. In humans, pre-term labour is defined as spontaneous labour occurring before 37 weeks of gestation (with 39 weeks being term). The usual calculated time of delivery for females as defined by the invention will be well known in the arts of
30 human and veterinary medicine.

Preferably, the medicament reduces and/or prevents fetal or neonatal damage.

More preferably, the medicament reduces and/or prevents one or more of
5 the following conditions: astrogliosis; loss of myelin-producing oligodendrocytes; multifocal necroses resulting in cystic change (periventricular leucomalacia, PVL).

By "astrogliosis" we include the meaning of hypertrophy (i.e. increasing
10 cell size) of the astroglia, that usually occurs in response to injury. Astroglia are the largest and most numerous neuroglial cells in the brain and spinal cord. Astrocytes (from "star" cells) are irregularly shaped with many long processes, including those with "end feet" which form the glial (limiting) membrane and directly and indirectly contribute to the blood-
15 brain barrier. They regulate the extracellular ionic and chemical environment, and "reactive astrocytes" (along with microglia) respond to injury. Astrocytes can release neuro-transmitters, but their role in signaling (as in many other functions) is not well understood.

20 By "oligodendrocytes" we include the meaning of neuroglial cell of the central nervous system (CNS) in vertebrates whose function is to myelinate CNS axons. "Loss of myelin-producing oligodendrocytes" means that there a reduction in the number of these cells.

25 By "multifocal necroses" we include the meaning of death of tissue occurring at more than one site. By "cystic change" we include the meaning of the development of fluid filled spaces in the region where necrosis has taken place. By "periventricular leucomalacia" or "PVL" we include the meaning of damage to the periventricular cerebral white matter
30 which is seen following cytokine induced or hypoxia/ischemia induced necroses and which can go on to become cystic change.

A particularly preferred embodiment of the invention is the use of the cyclopentenone prostaglandin 15-deoxy- $\Delta^{12,14}$ -prostaglandin J₂ and/or prostaglandin A₁.

5

Alternatively, the cyclopentenone prostaglandin is provided in the form of a prodrug of 15-deoxy- $\Delta^{12,14}$ -prostaglandin J₂ and/or prostaglandin A₁.

It will be appreciated by those skilled in the art that certain metabolic precursors of cyclopentenone prostaglandins, may not possess pharmacological activity as such, but may, in certain instances, be administered to a patient and thereafter metabolised in the body to form compounds of the invention which are pharmacologically active. Such derivatives may therefore be described as “prodrugs”.

15

All prodrugs of the cyclopentenone prostaglandins, particularly those of 15-deoxy- $\Delta^{12,14}$ -prostaglandin J₂ and/or prostaglandin A₁, are included within the scope of the invention.

20 Preferably, the prodrug is PGD₂ (the precursor of 15-dPGJ₂) or PGE₁ (the precursor of PGA₁).

Preferably, the medicament further comprises a pharmaceutically acceptable excipient, diluent or carrier.

25

By “pharmaceutically acceptable” we mean that the carrier does not have a deleterious effect on the recipient. Typically, the carrier will be sterile and pyrogen free.

30 Preferably the medicament is in a form adapted for delivery by mouth, intravenous injection or intra-amniotic injection.

Preferably, the medicament is in a form which is compatible with the amniotic fluid. More preferably, the medicament is in a form which has substantially the same pH and/or osmotic tension as amniotic fluid.

5

The amniotic fluid has a distinct pH and a distinct osmotic tension. The amniotic fluid pH and osmotic tension are well known to, or can be readily measured by, the person skilled in the art.

- 10 Preferably, the medicament further comprises an agent for treating a female who has or is at risk of one or more of the following conditions: pre-term labour; pathogenic infection; cervical ripening, uterine contractions.

- 15 By an "agent for treating a female who has or is at risk of one or more of the following conditions: pre-term labour; pathogenic infection; cervical ripening, uterine contractions" we include corticosteroids, tocolytic agents and anti-inflammatory prostaglandins.

Preferably, the agent is a corticosteroid.

20

More preferably, the agent is capable of preventing and/or reducing respiratory distress syndrome.

- 25 One function of corticosteroids is to mature the fetal lung, which leads to an increase in surfactant production and therefore prevents or reduces the severity of neonatal respiratory problems

More preferably, the agent is selected from dexamethasone or betamethasone. Such agents are known to those skilled in the art.

- 30 Administration of such agents may be two doses of 12mg intra-muscular (IM), 12 or 24 hours apart.

Preferably, the agent is capable of delaying delivery.

More preferably, the agent capable of delaying delivery is selected from:

- 5 oxytocin receptor antagonists; calcium channel blockers; sympathomimetics; nitric oxide donors

Preferably, the agent is a tocolytic agent.

- 10 By “tocolytic” we include the meaning of a drug whose action is to stop uterine contractions.

More preferably, the tocolytic agent is selected from: oxytocin receptor antagonists, calcium channel blockers, sympathomimetics, nitric oxide

- 15 donors.

More preferably, the oxytocin receptor antagonist is Atosiban. More preferably, the calcium channel blocker is Nifedipine. More preferably, the sympathomimetic is Ritodrine. More preferably, the nitric oxide donor is

- 20 glyceryl trinitrate.

Preferably, the inflammatory response is mediated by NF κ B in uterine cells.

More preferably, the cyclopentenone prostaglandin is capable of inhibiting and/or reducing NF κ B activity by preventing and/or reducing NF κ B DNA-binding in uterine cells.

- 25

More preferably, the cyclopentenone prostaglandin is capable of inhibiting and/or reducing NF κ B activity by preventing and/or reducing NF κ B-mediated transcriptional regulation in uterine cells.

- 30

More preferably, the cyclopentenone prostaglandin is capable of inhibiting and/or reducing NF κ B activity by preventing and/or reducing NF κ B production in uterine cells.

- 5 A further aspect of the invention is to provide a pharmaceutical composition comprising a cyclopentenone prostaglandin and a pharmaceutically acceptable carrier or excipient, the cyclopentenone prostaglandin being present in an amount effective to prevent and/or reduce an inflammatory response in the reproductive system of a female.

10

A further aspect of the invention is a method of treating inflammation within the reproductive system of a female, the method comprising administering an effective amount of a medicament of the invention.

- 15 A further aspect of the invention is to provide a method for identifying a cyclopentenone prostaglandin for delaying the onset and/or preventing the continuation of labour in a female comprising the step of testing the cyclopentenone prostaglandin to determine if it is capable of inhibiting and/or reducing NF κ B activity in uterine cells in a PPAR- γ independent
20 manner.

By "NF κ B activity" we include the DNA-binding activity of NF κ B and/or NF κ B-mediated transcriptional regulation.

- 25 Testing a cyclopentenone prostaglandin to determine if it is capable of inhibiting and/or reducing NF κ B activity in uterine cells in a PPAR- γ independent manner can be performed by the methods described in Example 1, below. For example, whether a cyclopentenone prostaglandin is capable of inhibiting and/or reducing NF κ B activity in uterine cells in a
30 PPAR- γ independent manner can be determined by using the PPAR- γ inhibitor GW-9662, as shown in Figure 6, below.

By "PPAR- γ independent manner" we include the meaning that the activity of a cyclopentenone prostaglandin occurs without it binding to and/or activating the PPAR- γ receptor.

5

It will be understood that a cyclopentenone prostaglandin to determine if it is capable of inhibiting and/or reducing NF κ B activity in uterine cells may be tested *in vitro*, *in vivo* or *ex vivo*.

10 A further aspect of the present invention is to provide a method for making a pharmaceutical composition for use in delaying the onset and/or preventing the continuation of labour in a female comprising providing a cyclopentenone prostaglandin identified by the method of the present invention and combining it with a pharmaceutically acceptable carrier.

15

Preferred, non-limiting examples which embody certain aspects of the invention will now be described, with reference to the following figures:

Figure 1 : 15dPGJ₂ inhibition of NF- κ B DNA binding

20

Electro-mobility shift assay (EMSA) analysis of NF- κ B DNA binding in nuclear protein extracts from (A) myometrial cells, (B) L+ amnion cells, and (C) L- amnion cells treated with 15dPGJ₂ or vehicle for 2 h +/- IL-1 β stimulation (15 min). Consensus κ B probe used to assess NF- κ B DNA
25 binding, and consensus Oct-1 probe used as control.

Figure 2 : PPAR- γ protein expression

Western immunoblots of (A) nuclear and cytosolic protein extracts from
30 myometrial and amnion cells with or without 15 min IL-1 β stimulation, and

(B) nuclear extracts of myometrial cells treated with 15d-PGJ₂ +/- IL-1b.
Probing with antibody to PPAR γ .

Figure 3 : *PPAR- α protein expression*

5

Western immunoblots of nuclear and cytosolic protein extracts from myometrial and amnion cells with or without 15 min IL-1b stimulation. Probing with antibody to PPAR α .

10 **Figure 4 :** *PPAR- γ agonists do not inhibit NF κ B DNA binding*

EMSA analysis of nuclear protein extracts from myometrial cells treated with (A) troglitazone, (B) GW1929 or vehicle for 2 h +/- IL-1b stimulation (15 min). Consensus κ B probe used to assess NF- κ B DNA binding,
15 consensus Oct-1 probe used as control. For supershift analysis, extracts were preincubated with antibodies against p50 or p65.

Figure 5 : *Troglitazone and WY-14643 do not inhibit NF κ B DNA binding*

20 EMSA analysis of nuclear protein extracts from myometrial cells treated with (A) WY-14643 or vehicle, and (B) high doses of troglitazone, WY-14643 or vehicle for 2 h followed by IL-1b stimulation (15 min). Consensus κ B probe used to assess NF κ B DNA binding, consensus Oct-1 and Sp-1 probes used as controls.

25

Figure 6 : *PPAR- γ antagonist GW9662 does not alleviate 15dPGJ₂ inhibition of NF κ B DNA binding*

EMSA analysis of nuclear protein extracts from amnion cells treated with
30 15dPGJ₂ +/- GW9662 or vehicle for 2 h followed by IL-1 β stimulation (15

min). Consensus κ B probe used to assess NF κ B DNA binding, consensus Oct-1 probe used as control.

Figure 7 : *PGA₁ inhibition of NF κ B DNA binding*

5

EMSA analysis of nuclear protein extracts from (A) myometrial cells, and (B) amnion cells treated with PGA₁ or vehicle for 2 h followed by IL-1 β stimulation (15 min). Consensus κ B probe used to assess NF κ B DNA binding, consensus Oct-1 probe used as control. For supershift analysis, extracts were preincubated with antibodies against p50 or p65.

10

Figure 8 : *Effect of cyPGs and PPAR agonists on NF κ B transcriptional activity in amnion*

15 Amnion cells derived from L- or L+ placentas were transiently transfected with the NF κ B-dependent reporter construct κ B.BG.Luc, treated with 15dPGJ₂, PGA₁, troglitazone, WY-14643, or vehicle for 2 h, and then stimulated with IL-1 β (1 ng/ml) for 6 h. The mutated κ Bmut.Luc construct was used as a control to confirm NF κ B-mediated transactivation. Values are normalised for b-gal reporter activity.

20

Figure 9 : *15dPGJ₂ inhibition of NF κ B transcriptional activity in myometrium*

25 Myometrial cells were transiently transfected with the NF κ B-dependent reporter construct κ B.BG.Luc, treated with 15dPGJ₂ or vehicle for 2 h, +/- IL-1 β (1 ng/ml) for 6 h. The mutated κ Bmut.Luc construct was used as a control to confirm NF κ B-mediated transactivation. Values are normalised for b-gal reporter activity. (NS = nonstimulated).

30

Figure 10 : *Effect of PGA_1 and PPAR agonists on $NF\kappa B$ transcriptional activity in myometrium*

Myometrial cells were transiently transfected with the $NF\kappa B$ -dependent
5 reporter construct $\kappa B.BG.Luc$, treated with troglitazone, WY-14643, PGA_1
or vehicle for 2 h, +/- IL-1b (1 ng/ml) for 6 h. The mutated $\kappa Bmut.Luc$
construct was used as a control to confirm $NF\kappa B$ -mediated transactivation.
Values are normalised for CMV-Renilla reporter activity. (NS =
nonstimulated).

10 **Figure 11 :** *PPAR- γ agonist GW1929 does not inhibit $NF\kappa B$ transcriptional activity*

Myometrial cells were transiently transfected with the $NF\kappa B$ -dependent
15 reporter construct $\kappa B.BG.Luc$, treated with GW1929 or vehicle for 2 h, +/-
IL-1b (1 ng/ml) for 6 h. Values are normalised for CMV-Renilla reporter
activity. (NS = nonstimulated).

20 **Figure 12 :** *Troglitazone and GW1929 potentiate PPAR- γ activation of a PPRE reporter*

Myometrial cells were cotransfected with 0.4 mg of the PPAR- γ -dependent
reporter construct 3-PPRE-TK.pGL3 and 100 ng, 200 ng or 300 ng of a
PPAR- γ expression construct. Cells were treated with 10 mM or 20 mM of
25 (A) troglitazone or (B) GW1929, or vehicle for 24 h. Values are normalised
for CMV-renilla reporter activity. Similar results were obtained with
transfection of amnion cells.

30 **Figure 13 :** *Troglitazone does not inhibit $NF\kappa B$ transcriptional activity in PPAR- γ -transfected cells*

Myometrial cells were transfected with 0.4 mg κ B.BG.Luc reporter and 200 ng PPAR- γ expression vector and treated with 10 mM troglitazone or vehicle for 7 h +/- IL-1b (1 ng/ml) for 17 h. Values are normalised for
5 CMV-renilla reporter activity. (*NS* = nonstimulated).

Figure 14 : *PPAR- γ overexpression does not potentiate 15d-PGJ₂ inhibition of NF κ B activity*

10 Myometrial cells were transfected with 0.4 mg κ B.BG.Luc reporter and 200 ng PPAR- γ expression vector and treated with 15d-PGJ₂ for 2h followed by IL-1 β (1 ng/ml) for 6 h. Values are normalised for β -galactosidase reporter activity. (*NS* = nonstimulated).

15 **Figure 15 :** *15dPGJ₂ inhibition of p65 nuclear localisation, p50 phosphorylation, and I κ B α degradation*

Western immunoblots of nuclear or cytosolic protein extracts from (A) myometrial cells, (B) L- amnion cells, and (C) L+ amnion cells treated with
20 15dPGJ₂ or vehicle for 2 h +/- IL-1 β stimulation (15 min). Blots probed with antibodies against p65, p50 or I κ B α .

Figure 16 : *PGA₁ inhibition of p65 nuclear localisation and I κ B α degradation*

25

Western immunoblot of nuclear or cytosolic protein extracts from myometrial cells treated with PGA₁ or vehicle for 2h followed by IL-1 β stimulation (15 min). Blots probed with antibodies against p65 or I κ B α .

Figure 17 : *PGE₂ does not inhibit TNF α - and IL-1 β -induced NF κ B activation*

5 Analysis of nuclear protein extracts from myometrial cells treated with PGE₂ or vehicle for 2 h +/- TNF α or IL-1 β stimulation (15 min). (A) EMSA using consensus κ B probe. (B) Western immunoblot probing for nuclear p65.

Figure 18 : *PGE₂ does not induce NF κ B DNA binding*

10 EMSA analysis of nuclear protein extracts from (A) L- amnion cells and (B) myometrial cells treated with vehicle, PGE₂ or IL-1 β . Consensus κ B probe used.

15 **Figure 19 :** *15dPGJ₂ inhibits I κ B α phosphorylation*

Western immunoblots of cytosolic extracts from myometrial cells (A) treated with 15dPGJ₂ for 2h +/- IL1 β for 15min; probed for IKK, and (B) treated with 30mM 15dPGJ₂, 40mM MG132 or vehicle for 2h, +/- IL-1 β for 20 15min; probed for I κ B α .

Figure 20 : *Effect of 15dPGJ₂ and PPAR agonists on IL-1 β -induced COX-2 protein expression*

25 Western immunoblot of cytosolic protein extracts from myometrial cells treated with 15dPGJ₂, troglitazone, WY-14643 or vehicle for 2h, followed by IL-1 β stimulation for 6h. Probed with antibodies to (A) COX-2, and (B) a smooth muscle actin.

Figure 21 : *Schematic of the structure of (A) prostaglandin A₁ (PGA₁) and (B) 15-deoxy- $\Delta^{12,14}$ prostaglandin J₂ (15-dPGJ₂)*

Figure 22 : *Effect of LPS and 15d-PGJ₂ on inflammatory responses – IL-1 β levels*

Concentrations of IL-1 β in placental homogenates collected from gestation day 16 mice 6 hours after intrauterine injection of 250 μ g LPS + vehicle or 250 μ g LPS + 4 μ g 15d-PGJ₂. * denotes statistically significant difference (t-test (p<0.05)).

Figure 23 : *Effect of LPS and 15d-PGJ₂ on inflammatory responses – phospho-p65 levels*

Relative concentrations of phospho-p65 in placental homogenates collected from gestation day 16 mice 6 hours after intrauterine injection of 250 μ g LPS + vehicle or 250 μ g LPS + 4 μ g 15d-PGJ₂. * denotes statistically significant difference (t-test (p<0.05)).

Figure 24 : *The cyclopentenone ring is essential for cyPG inhibition of NF- κ B.*

(A) NF- κ B-DNA binding was measured by EMSA in nuclear protein extracts from myometrial cells pre-treated with vehicle, 15d-PGJ₂ or PGA₁ for 2 h, followed by stimulation with IL-1 β (1 ng/ml) for 15 min. Antibodies against p50 and p65 were used for supershift analysis. (B) Myometrial cells were transiently transfected with a NF- κ B-LUC reporter and a β -gal reporter plasmid, pre-treated with vehicle or PGA₁ for 2 h, and stimulated with IL-1 β (1 ng/ml) for 6 h. Luciferase activity was normalized for β -gal reporter readout. Values are presented as the mean \pm SEM obtained for each treatment done in triplicate. Western blot analysis of

nuclear p65 and p50 expression in myometrial cells treated with (C) PGA₁ or (D) 15d-PGJ₂ for 2 h, followed by stimulation with IL-1 β (1 ng/ml) for 15 min. (E) Western blot analysis of whole cell lysates from myometrial cells treated with 15d-PGJ₂ or 9,10-dihydro-15d-PGJ₂ for 2 h, followed by
 5 stimulation with IL-1 β (1 ng/ml) for 15 min. Membranes were probed with antibodies against p65 and Ser 536-phosphorylated p65. Similar results were obtained in amnion epithelial cells.

EXAMPLE 1 – Experimental data

10

Methods

Abbreviations

EDTA	Ethylenediaminetetraacetic acid
EGTA	Ethyleneglycol bis-aminoethyltetra acetic acid
DTT	Dithiotreitol
HEPES	4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid
NP-40	Nonidet P-40
SDS-PAGE	Sodium dodecyl sulphate – Polyacrylamide gel electrophoresis
PVDF	Polyvinylidene difluoride
PBS-T	Phosphate Buffered Saline plus Tween
HRP	Horseradish peroxidase
PBS	Phosphate Buffered Saline
FCS	Foetal Calf Serum
DMEM	Dulbecco's modified eagle's medium

15

Tissue biopsies and cell culture

Local Ethics committee approval was obtained for the collection of these tissues and patients gave informed consent.

20

Human myometrial cell culture

Myometrial tissue was collected at term from the upper margin of uterine

incision at the time of lower segment caesarean section either prior to the onset of labour (L-) or during fetal distress (L+). L+ samples were collected by Dr Mark Johnson and Dr S Soorrana at Chelsea & Westminster Hospital. Myometrial tissue was dissected, rinsed in PBS, and digested in
5 serum-free DMEM containing 15mg/ml collagenase 1A (Sigma), 15mg/ml collagenase X, and 50mg/ml bovine serum albumin for 45min at 37°C. The cell suspension was filtered through a cell strainer, centrifuged at 400g for 5min, and the pellet re-suspended and plated out in DMEM, 10% FCS (Helena BioScience), 1% L-glutamine, 1% penicillin-streptomycin. Cells
10 were used between passage numbers 1-4.

Human Amnion Cell Culture

Placentae were obtained from patients at term either at elective Caesarean
15 section prior to labour (L-) or following spontaneous labour onset and vaginal delivery (L+). Amnion cells were prepared as described in Bennett *et al.*, (1989). Briefly, the amnion was separated from the placenta, washed 3x in PBS, cut into strips, and incubated in 0.5mM EDTA in PBS for 15min. The strips were washed in PBS 2x and digested with 2.5mg/ml
20 dispase in serum-free DMEM for 35min at 37°C. The amnion was then shaken vigorously in DMEM, 10% FCS to dissociate the cells, the remaining strips discarded, and the cell suspension pelleted at 175g for 10min and cultured in DMEM, 10% FCS (Sigma), 1% L-glutamine, 1% penicillin-streptomycin.

25

Protein Extracts from cultured cells

Nuclear and cytosolic protein extracts were obtained from cultured amnion cells as described by Schreiber *et al* (1989). For nuclear/cytosolic
30 fractionation, confluent cell monolayers were scraped and lysed using a buffer containing 10mM HEPES, 10mM KCl, 0.1mM EDTA, 0.1mM

EGTA, 2mM DTT, 1% (v/v) NP-40 and complete protease inhibitor tablets (CPIs, Roche), diluted to manufacturer's instructions. Cell lysates were incubated on ice for 10min and NP-40 added to a final concentration of 1% (v/v). Lysates were vortexed for 10secs and centrifuged for 30secs at 4°C,
5 12000g. The supernatants were retained as the cytosolic protein extracts. The pellets were resuspended in buffer containing 10mM HEPES, 10mM KCl, 0.1mM EDTA, 0.1mM EGTA, 2mM DTT, 400mM NaCl, 1% NP-40 (v/v) and CPIs. Samples were shaken vigorously for 15min in an ice bath. The nuclear protein extracts were obtained in the supernatant following a
10 5min centrifugation at 4°C, 12000g.

For whole cell lysates, confluent cell monolayers were scraped and lysed in a high-salt extraction buffer containing 0.4M KCl, 20mM HEPES, 20% (v/v) glycerol, 1mM DTT, and CPIs.

15

Protein Extracts from fresh tissue biopsies

Tissue samples were rinsed in ice-cold PBS, dissected, flattened between aluminium foil, flash-frozen in liquid nitrogen, and stored at -80°C.
20 Samples were reduced to powder in liquid nitrogen using a pestle and mortar. Powdered tissue was homogenized in a Dounce homogeniser on ice in a buffer containing 0.6% (v/v) NP-40, 150mM HEPES, 1mM EDTA, 0.5mM PMSF and any unbroken tissue was removed by centrifugation for 30sec at 2000rpm at 0°C. The supernatant was incubated on ice for 5min,
25 centrifuged for 10min at 4000rpm at 0°C, and the nuclear pellets resuspended in 25% (v/v) glycerol 20mM HEPES, 0.42M NaCl, 1.2 mM MgCl₂, 0.2mM EDTA, 0.5mM DTT, and CPIs.

All extracts were aliquoted, frozen on dry ice and stored at -80°C. The
30 extracts were processed for protein quantitation by the Lowry method using

Bio-Rad protein assay reagents (Bio-Rad Laboratories) according to manufacturer's instructions.

Electro-mobility shift assay (EMSA)

5

Oligonucleotide labelling

Sense and antisense strands (175nmole/ml each) were incubated in annealing buffer (10mM Tris-HCl pH7.5, 100mM NaCl, 1mM EDTA) for
 10 10min at 65°C, and allowed to cool at room temperature for 2h. 3.5 pmole double-stranded oligonucleotides were end-labelled with 0.37MBq ³²P(γATP) by incubating for 30min at 37°C with T4 polynucleotide kinase. Labelled oligonucleotides were recovered by centrifugation at 3000rpm for
 15 2min through MicroSpin G-25 or G-50 sephadex columns (Amersham Biosciences).

EMSA

3-5μg protein extracts were incubated on ice for 1h with non-radiolabelled
 20 non-specific oligonucleotide (poly(dI-dC) or Oct-1) in a binding buffer (20% (v/v) glycerol, 5mM MgCl₂, 2mM EDTA, 50mM Tris-HCl pH7.5, 250mM NaCl, 2mM DTT), followed by a 45min incubation with 0.035pmole ³²P(γATP)-end labelled oligonucleotide probes:

25 consensus NF-κB: 5'-AGT TGA GGG GAC TTT CCC AGG C-3'
 consensus Oct-1: 5'-TGT CGA ATG CAA ATC ACT AGA A-3'
 consensus SP-1: 5'-ATT CGA TCG GGG CGG GGC GAG
 upstream COX-2 κB: 5'-CGG GAG AGG GGA TTC CCT GCG C-3'
 downstream COX-2 κB: 5'-AGA GTG GGG ACT ACC CCC TCT-3'

30

Oct-1 or SP-1 consensus sequences were used as a controls for a NF- κ B-specific effect. The resulting protein/DNA complexes were separated in a 4% acrylamide gel, the gel dried under vacuum at 80°C and exposed to X-ray film. For supershift analysis, samples were incubated with 2 μ g antibodies for 30min on ice prior to incubation with oligonucleotides. Non-radio-labelled oligonucleotides were used at 100-fold molar excess for specific and non-specific competition for DNA binding. Reagents for EMSA were obtained from Promega Life Sciences, Delta House, Chilworth Research Centre, Southampton SO16 7NS, United Kingdom.

SDS-PAGE and Western blotting analysis

Protein samples (20-70 μ g) were mixed with Laemmli sample buffer (1:1) containing β -mercaptoethanol (5%), and boiled for 5min. Proteins were then separated by SDS-PAGE (12-14% gels) and transferred onto PVDF membrane (Amersham Pharmacia Biotech). The membranes were blocked overnight in 5% non-fat milk prepared in PBS-T buffer, at 4°C. The blots were incubated with the primary antibody in 1% non-fat milk in PBS-T buffer for 1h, and washed three times (10min each) in PBS-T with vigorous shaking. The blots were then incubated with HRP-conjugated secondary antibody (diluted 1:2000 in 1% non-fat milk in PBS-T buffer) for 1h and washed three times (10min each) in PBS-T. Signal detection was achieved using enhanced chemi-luminescence (ECL plus system, Amersham Pharmacia Biotech) according to manufacturer's instructions.

To re-probe a membrane, blots were incubated for 30min in 50°C stripping buffer (2% SDS, 62.5mM Tris-HCl pH6.7, 100mM 2-mercaptoethanol), washed 2x in PBS-T, placed in blotto overnight, and then probed with a new antibody as above.

30-50µg protein extracts were subjected to SDS-PAGE and Western immuno-blotting. Secondary antibodies were IgG-HRP and ECL Plus detection kit (Amersham Pharmacia Biotech, Amersham Place, Little Chalfont, Bucks, HP7 9NA) was used for visualisation.

5

Transfections and luciferase assay

Cells at 70-80% confluence in 24-well plates were transfected using the liposome Transfast (Promega). 0.5µg per well of luciferase reporter
10 construct was transfected using a 1:1 ratio of transfection (i.e., 3µl Transfast per 1µg DNA) in serum-free DMEM for 1h. DMEM, 10% FCS was then added and the cells were incubated at 37°C for 24h. The medium was replaced with DMEM, 2% FCS for a further 24h, and the cells treated with various agonists/inhibitors or vehicle for 6-8h. Transfections were analysed
15 in a dual firefly/renilla (Packard BioSciences/Calbiochem) luciferase assay or firefly/β-galactosidase (Promega/Galacton) assay using a luminometer.

pGL3.6κB.BG.luc was the reporter construct used to assess NF-κB-mediated transcription, while the mutant pGL3.6κBmut.luc and empty
20 pGL3.BG.luc were used as controls (Schwarzer *et al.*, 1998).

pGL3.6κB.BG.luc: a NF-κB-dependent reporter construct with 6 copies of the NF-κB binding site. It contains two tandem repeats of the sequence 5'-
GGG GAC TTT C CC TGG GGA CTT TCC CTG GGG ACT TTC CC-3',
25 which contains three copies of the decameric NF-κB binding site (underlined) upstream of a minimal β-globin promoter driving a luciferase gene.

pGL3.6κBmut.luc: this reporter construct is as above except that the core NF-κB binding site is mutated to 5'-GCC ACT TTC C-3' (mutated bases underlined).

- 5 pGL3.BG.luc: this reporter construct contains only the minimal β-globin promoter.

Cells were co-transfected with the renilla vector pRL-CMV or a β-galactosidase vector pCH110 as internal controls for transfection
10 efficiencies.

In vitro translation and plasmid preps

For recombinant production of p65, a pSG5/p65 expression construct was
15 transcribed and translated using a TNT Coupled Reticulocyte Lysate System (Promega), according to manufacturer's instructions. QIAGEN Maxi Prep kits were used for plasmid isolation from transformed JM109 *E. coli* cells, and all constructs were subsequently precipitated with polyethylene glycol.

20

Reagents/Antibodies

Recombinant cytokine IL-1β and TNFα from R&D Systems; 15d-PGJ₂, PGA₁, troglitazone, GW-9662, and 16,16-dimethyl-PGE₂ from Cayman
25 Chemical; WY-14643, MG132 proteasome inhibitor, and PG490 (triptolide) from Calbiochem; HRP-conjugated secondary antibodies and antibodies to p50, p65, c-rel, Rel B, COX-2, IκBα, IκBβ, and PPARγ from Santa Cruz; antibodies to p52, Bcl-3 and smooth muscle actin from Upstate Biotechnologies. Antibody to PPAR-γ from Affinity BioReagents, to
30 phospho-p65 from Cell Signaling, to COX-1 from Alexis Biochemicals, and to lamin B from Oncogene Research Products.

Mouse model of preterm labour

Surgery was performed on timed pregnant MF1 mice at day 16 of gestation.
5 After deep maternal anaesthesia was attained, a minilaparotomy was performed in the lower abdomen. The uterine horns were exposed through the incision and preterm labour was induced by the intrauterine injection of 250µg lipopolysaccharide (LPS, Sigma) into the gravid horn. This was immediately followed by injection of 4µg 5d-PGJ₂ (Cayman), or an equal
10 volume of vehicle (methyl acetate), at the same site. The uterus was then returned to the abdomen and the fascia and skin were closed with continuous vicryl sutures.

Effect of LPS and 15d-PGJ₂ on inflammatory responses

15 Mice were sacrificed 6 hours after injection of LPS ± 15d-PGJ₂. Placentae were washed in phosphate buffered saline (PBS), flash frozen in liquid nitrogen and stored at -80°C until further processing. Fetuses were washed in PBS, then immediately fixed in 4% paraformaldehyde for 24h and then
20 stored in 70% ethanol until further processing. Placentae were homogenized for 1 minute in the presence of lysis buffer comprising 400mM KCl, 20mM HEPES pH7.4, 1mM dithiothreitol, 20% glycerol and 5% (v/v) protease inhibitor cocktail.

25 Homogenate levels of Interleukin-1β (IL-1β) and tumour necrosis factor α (TNFα) were determined in placental lysates by ELISA (R and D systems) according to manufacturers instructions. Total protein concentrations were determined for each homogenate and IL-1β and TNFα levels were expressed as pg/mg total protein. Homogenates were also subjected to
30 polyacrylamide gel electrophoresis. Loading volumes were adjusted according to the protein content of each homogenate such that a constant

amount of protein was run in each lane. Phosphorylated p65 (phospho-p65) was detected by western immunoblotting using a specific antibody (Santa Cruz) and quantified by densitometric analysis.

5

Results

CyPGs, but not PPAR agonists, inhibit NF- κ B DNA binding in amnion and myometrial cells.

15d-PGJ₂ inhibited IL1- β -induced NF- κ B DNA binding in a dose-dependent manner in myometrial cells, as well as in L- and L+ amnion cells (*Fig. 1*). Protein binding to a consensus Oct-1 or Sp-1 probe was unaffected by either IL-1 β or 15d-PGJ₂ treatment, confirming that the effects observed are NF- κ B-specific.

Since PPAR- γ is the putative endogenous receptor for 15d-PGJ₂, and PPAR expression may be affected by cytokines (Tontonoz *et al.*, 1998, Tanaka *et al.*, 1999), PPAR- γ protein expression was examined in myometrial and amnion cells. PPAR- γ was shown to be expressed predominantly in the nucleus of both cell types, and its expression was not affected by IL-1 β or 15d-PGJ₂ treatment (*Fig. 2*). 15d-PGJ₂ can also transactivate PPAR- α , though more weakly than PPAR- γ (Forman *et al.*, 1995). PPAR- α expression in myometrial and amnion cells was found to be predominantly cytoplasmic (*Fig. 3*).

The ability of synthetic PPAR agonists to mimic the inhibitory effects of 15d-PGJ₂ was examined. The PPAR- γ agonist troglitazone had no effect on NF- κ B DNA binding at 10-50 μ M doses, although it did cause a slight reduction at 100 μ M (*Fig. 4, 5*). Troglitazone can transactivate PPAR- γ at

1 μ M and induces weak interactions between PPAR- γ and the co-activators
p300 and steroid receptor co-activator (SRC-1) at 10 μ M doses;
adipogenesis is positively regulated by PPAR- γ , and troglitazone can induce
expression of adipogenic markers at 5-10 μ M doses (Prusty *et al.*, 2002).

5 Thus, at 100 μ M concentrations, it is unlikely that troglitazone is exerting a
specific effect through PPAR- γ . Since structurally distinct PPAR ligands
may differentially affect coactivator/corepressor recruitment, a new potent
PPAR- γ agonist, which lacks the TZD moiety, was also used. This
GW1929 ligand failed to inhibit NF- κ B DNA binding (*Fig. 6*). The
10 synthetic PPAR- α agonist WY-14643 can transactivate PPAR α at 5-25 μ M
doses in a GAL4 chimera transfection system (Kehrer *et al.*, 2001), but
WY-14643 had no effect on NF- κ B DNA binding, at 10-100 μ M
concentrations. To further investigate a potential role for PPAR- γ in
mediating the inhibitory effects of 15d-PGJ₂, NF- κ B DNA binding was
15 assessed in cells treated with 15d-PGJ₂ in the presence of the selective
PPAR- γ inhibitor GW-9662. GW9662 binds irreversibly to PPAR- γ through
covalent modification of Cys²⁸⁵ in the ligand-binding domain (Leesnitzer *et al.*, 2002). GW-9662 failed to alleviate 15d-PGJ₂ inhibition of NF- κ B (*Fig.*
6).

20

In contrast, PGA₁, which does not act as a PPAR ligand but does contain a
cyclopentenone ring, was able to inhibit NF- κ B DNA binding in amnion
and myometrial cells, albeit at much higher doses than 15d-PGJ₂ (*Fig. 7*).

25 *CyPGs, but not PPAR agonists, inhibit NF- κ B transcriptional activity.*

To determine whether the cyPG effects on NF- κ B DNA binding extend to
inhibition of NF- κ B transactivation potential, amnion cells were transfected
with the NF- κ B-dependent reporter κ B.BG.Luc and treated with 15d-PGJ₂,
30 PGA₁, troglitazone, WY-14643 or vehicle, followed by IL-1 β stimulation

(Fig. 8). Constitutive reporter activity was seen in both L- and L+ amnion cells, although the levels were lower and showed a greater increase with IL-1 β in L- cells, in agreement with previous studies by Allport *et al* (2001). Both 15d-PGJ₂ and PGA₁ inhibited IL-1 β -induced NF- κ B transcriptional activity, whereas troglitazone and WY-14643 did not.

In myometrial cells, 15d-PGJ₂ inhibited IL-1 β -induced NF- κ B transcriptional activity in a dose-dependent manner, reducing reporter activity to basal levels (Fig. 9). IL-1 β -induced NF- κ B transcriptional activity was also reduced to basal levels by PGA₁, but not troglitazone, GW1929 or WY-14643 (Fig. 10, 11).

GW1929 and troglitazone were shown to be functional as PPAR- γ ligands, potentiating PPAR- γ -mediated transcription of a PPRE-dependent reporter in both cell types. Endogenous PPAR- γ levels were not sufficient to drive the PPRE reporter in the transfection system used, with transcription requiring overexpression of the receptor. Troglitazone was also unable to inhibit a NF- κ B-dependent reporter in PPAR γ -transfected cells, and PPAR γ overexpression did not promote 15d-PGJ₂ inhibition of NF- κ B transcriptional activity (Fig. 12, 13, 14).

CyPGs, but not PGE₂, inhibit NF- κ B activation and I κ B degradation.

15d-PGJ₂ inhibited IL-1 β -induced p65 nuclear translocation and p50 phosphorylation in myometrial cells and in L-, L+ amnion cells in a dose-dependent manner (Fig. 15). This was paralleled by inhibition of IL-1 β -induced I κ B α and I κ B β degradation. Similarly, PGA₁ inhibited p65 nuclear translocation and I κ B α degradation in myometrial cells (Fig. 16).

16,16-Dimethyl-PGE₂, a PGE₂ analogue with increased half-life, did not inhibit NF-κB DNA binding (controlled for with Oct-1 binding) or IL-1β-induced p65 nuclear translocation in myometrial and amnion cells (*Fig. 17*). This is not unexpected, since, in contrast to the cyPGs, PGE₂ is known to be
5 pro-inflammatory, does not contain a cyclopentenone ring, and does not activate PPAR-γ (Forman *et al.*, 1995). 16,16-dimethyl-PGE₂ did not inhibit NF-κB DNA binding or p65 nuclear translocation in myometrial cells (*Fig. 18*). However, neither did it stimulate NF-κB activity as reported in T cells (Dumais *et al.*, 1998), nor did it synergise with IL-1β or TNFα.

10

Effect of 15-dPGJ₂ on NF-κB upstream activators and downstream targets.

In contrast to the proteasome inhibitor MG132, which prevented IL-1β-induced IκBα degradation and resulted in the accumulation of undegraded,
15 phosphorylated IκBα, accumulation of phosphorylated IκBα was not detected following 15-dPGJ₂ treatment, suggesting that 15-dPGJ₂ may be affecting IKKs or other upstream kinases (*Fig. 19*). Both IL-1β and 15-dPGJ₂ treatment had no effect on IKKα or IKKβ protein expression, although it is more likely that 15d-PGJ₂ would inhibit the kinase activity of
20 the IKKs.

Since COX-2 is an important target gene for NF-κB in labour, the effect of 15-dPGJ₂ and PPAR agonists on COX-2 expression was assessed. IL-1β-induced COX-2, expression was inhibited by 15-dPGJ₂, but not by
25 troglitazone or WY-14643 (*Fig. 20*). Similar results were obtained in L- and L+ amnion cells.

Effect of LPS on pre-term delivery.

Pre-term delivery of pups occurred by 16 hours after injection of LPS using the mouse model of preterm labour, as set out in the methods above.

Effect of LPS and 15d-PGJ₂ on inflammatory responses

5

In all mice, levels of TNF α and IL-1 β were significantly higher in the placenta proximal to the injection site compared to those in the opposite horn. Levels of IL-1 β were approximately 40% lower in proximal placenta injected with LPS + 15d-PGJ₂ compared to those given LPS + vehicle (Fig. 10 22). This difference was statistically significant ($p < 0.05$). In contrast, TNF α levels were not significantly altered according to drug treatment.

Significantly, placental levels of IL-1 β were not altered according the proximity of the placenta to the site of injection, indicating that 15 inflammatory response can be distributed throughout the uterus, irrespective of the site of infection. However, phospho-p65 levels were approximately 35% lower in proximal placenta injected with LPS + 15d-PGJ₂ compared to those given LPS + vehicle (Fig. 23) and this difference was statistically significant ($p < 0.05$).

20

The cyclopentenone ring is essential for cyPG inhibition of NF- κ B

Several studies have demonstrated that 15d-PGJ₂ is a PPAR agonist, whilst other prostaglandins, such as PGA₁, are not (Forman *et al.*, 1995; Kliewer *et al.*, 1995; Ferry *et al.*, 2001). We have shown that PGA₁ shares the effect of 25 15d-PGJ₂ on NF- κ B, but that 9,10-dihydro-15d-PGJ₂ (an analogue of 15d-PGJ₂ which retains PPAR γ agonist activity but in which the cyclopentenone ring has been disrupted) could not reproduce the effects of 15d-PGJ₂ (Fig. 24). Taken together, these findings indicate that the inhibitory effects of 30 15d-PGJ₂ on NF- κ B in amnion epithelial and myometrial cells can be

attributed to its electrophilic ring, that similar effects would be expected with other cyclopentenone prostaglandins but not with other, non-cyclopentenone PPAR agonists.

5 *Conclusions*

NF-κB inhibition by cyPGs

15d-PGJ₂ inhibited IL1-β-induced NF-κB DNA binding and NF-κB-
10 mediated transactivation in myometrial cells, as well as in L- and L+ amnion cells. 15d-PGJ₂ inhibited the nuclear translocation and activation of NF-κB, at least in part, by preventing the degradation of IκBα by IL-1β.

In myometrial and amnion cells, which expressed both PPAR-α and PPAR-
15 γ receptors, neither PPAR-γ nor PPAR-α agonists were able to inhibit IL-1β-induced NF-κB DNA binding or NF-κB transcriptional activity at doses shown to inhibit NF-κB in other cell types (Chinetti *et al.*, 1998; Gupta *et al.*, 2001), or even at higher concentrations. In a study investigating the potential functional interactions between PPAR-γ and NF-κB in adipocytes,
20 PPAR-γ agonists did not impair TNFα-induced NF-κB activation, nuclear translocation, or DNA binding activity; rather, they antagonised the transcriptional regulatory activity of NF-κB, and PPAR-γ overexpression was required to demonstrate such inhibition (Ruan *et al.*, 2003). In the present study, while PPAR-γ overexpression potentiated transactivation of a
25 PPRE, it did not enable the PPAR-γ agonists to inhibit NF-κB transcription. In addition, 15d-PGJ₂ was able to inhibit NF-κB transcription in the absence of exogenous PPAR-γ and overexpression of this receptor did not promote inhibition.

IL-1 β -induced COX-2 expression was inhibited by 15d-PGJ₂ but not by PPAR agonists. While PPAR agonists are known to be anti-inflammatory and can inhibit COX-2 expression (Stael *et al.*, 1998; Subbaramaiah *et al.*, 2001), they have also been reported to enhance COX-2 expression in certain
5 cell types (Meade *et al.*, 1999; Ikawa *et al.*, 2001; Pang *et al.*, 2003).

CyPGs such as 15d-PGJ₂ are characterised by the presence of a cyclopentenone ring system containing an electrophilic carbon. This ring can react covalently with nucleophiles such as the free sulfhydryls of
10 glutathione and cysteine residues in cellular proteins. Receptor-independent actions of 15d-PGJ₂ have been attributed to its cyclopentenone ring. NF- κ B proteins contain a conserved cysteine residue in their DNA-binding domain (DBD) and alkylation of this cysteine impairs DNA binding (Toledano *et al.*, 1993). In the present study, PGA₁, a cyPG that does not act as a PPAR-
15 γ ligand, was able to inhibit NF- κ B DNA binding and transactivation, albeit at higher concentrations than 15d-PGJ₂. This ability of PGA₁, but not PGE₂ or PPAR agonists, to mimic the effects of 15d-PGJ₂ suggests that these cyPGs may inhibit NF- κ B in amnion and myometrial cells by virtue of their cyclopentenone ring. Indeed, our results indicate that the inhibitory effects
20 of 15d-PGJ₂ on NF κ B in amnion epithelial and myometrial cells can be attributed to its electrophilic ring and that similar effects would be expected with other cyclopentenone prostaglandins but not with other, non-cyclopentenone PPAR agonists.

25 While direct modification of NF- κ B cysteines has not been addressed in this study, both 15d-PGJ₂- and PGA₁-mediated inhibition of NF- κ B was shown to involve the inhibition of I κ B α degradation, suggesting that events further upstream in the NF- κ B cascade are being targeted.

Thus, while PPAR activation may not be effectively anti-inflammatory in amnion and myometrium, the use of cyPGs should prove useful in repressing NF- κ B, and therefore an array of pro-inflammatory and labour-associated genes, in these tissues. CyPG administration offers an attractive
5 alternative approach to anti-inflammatory treatment since a potential specificity of cyPGs for IKK β /I κ B α would spare other potentially beneficial pathways of NF- κ B activation (e.g., the processing of p105 and formation of p50 homodimers), which might be disrupted by more broad-spectrum NF- κ B inhibitors. The use of the cyPGs, able to simultaneously
10 trigger the inhibition of the pro-inflammatory NF- κ B and harness the anti-inflammatory activities of endogenous cytoprotective molecules represents a novel therapeutic approach in the treatment of preterm labour and neurodevelopmental disorders of the neonate.

15 This study provides evidence that the mouse model used is an effective model for the study of preterm delivery and agents that may delay the onset of preterm delivery. The finding of lower levels of IL-1 β and phospho-p65 in mice treated with the cyclopentenone prostaglandin 15d-PGJ₂ suggests that this compound is effective at blocking the inflammatory pathway
20 induced by LPS treatment *in vivo*.

EXAMPLE 2 – Preferred pharmaceutical formulations and modes and doses of administration.

5 The compounds of the present invention may be delivered using an injectable sustained-release drug delivery system. These are designed specifically to reduce the frequency of injections. An example of such a system is Nutropin Depot which encapsulates recombinant human growth hormone (rhGH) in biodegradable microspheres that, once injected, release rhGH slowly over a sustained period.

10

The compounds of the present invention can be administered by a surgically implanted device that releases the drug directly to the required site. For example, Vitrasert releases ganciclovir directly into the eye to treat CMV retinitis. The direct application of this toxic agent to the site of disease
15 achieves effective therapy without the drug's significant systemic side-effects.

Electroporation therapy (EPT) systems can also be employed for administration. A device which delivers a pulsed electric field to cells
20 increases the permeability of the cell membranes to the drug, resulting in a significant enhancement of intracellular drug delivery.

Compounds can also be delivered by electroincorporation (EI). EI occurs when small particles of up to 30 microns in diameter on the surface of the
25 skin experience electrical pulses identical or similar to those used in electroporation. In EI, these particles are driven through the stratum corneum and into deeper layers of the skin. The particles can be loaded or coated with drugs or genes or can simply act as "bullets" that generate pores in the skin through which the drugs can enter.

30

An alternative method of administration is the ReGel injectable system that is thermosensitive. Below body temperature, ReGel is an injectable liquid while at body temperature it immediately forms a gel reservoir that slowly erodes and dissolves into known, safe, biodegradable polymers. The active
5 drug is delivered over time as the biopolymers dissolve.

The compounds of the invention can also be delivered orally. The process employs a natural process for oral uptake of vitamin B₁₂ in the body to co-deliver proteins and peptides. By riding the vitamin B₁₂ uptake system, the
10 protein or peptide can move through the intestinal wall. Complexes are synthesised between vitamin B₁₂ analogues and the drug that retain both significant affinity for intrinsic factor (IF) in the vitamin B₁₂ portion of the complex and significant bioactivity of the drug portion of the complex.

15 Compounds can be introduced to cells by "Trojan peptides". These are a class of polypeptides called penetratins which have translocating properties and are capable of carrying hydrophilic compounds across the plasma membrane. This system allows direct targeting of oligopeptides to the cytoplasm and nucleus, and may be non-cell type specific and highly
20 efficient (Derossi *et al.*, 1998).

Preferably, the pharmaceutical formulation of the present invention is a unit dosage containing a daily dose or unit, daily sub-dose or an appropriate fraction thereof, of the active ingredient.

25

The compounds of the invention can be administered orally or by any parenteral route, in the form of a pharmaceutical formulation comprising the active ingredient, optionally in the form of a non-toxic organic, or inorganic, acid, or base, addition salt, in a pharmaceutically acceptable
30 dosage form. Depending upon the disorder and patient to be treated, as well

as the route of administration, the compositions may be administered at varying doses.

Formulations in accordance with the present invention suitable for oral
5 administration may be presented as discrete units such as capsules, cachets
or tablets, each containing a predetermined amount of the active ingredient;
as a powder or granules; as a solution or a suspension in an aqueous liquid
or a non-aqueous liquid; or as an oil-in-water liquid emulsion or a water-in-
oil liquid emulsion. The active ingredient may also be presented as a bolus,
10 electuary or paste.

A tablet may be made by compression or moulding, optionally with one or
more accessory ingredients. Compressed tablets may be prepared by
compressing in a suitable machine the active ingredient in a free-flowing
15 form such as a powder or granules, optionally mixed with a binder
(*e.g.* povidone, gelatin, hydroxypropylmethyl cellulose), lubricant, inert
diluent, preservative, disintegrant (*e.g.* sodium starch glycolate, cross-linked
povidone, cross-linked sodium carboxymethyl cellulose), surface-active or
dispersing agent. Moulded tablets may be made by moulding in a suitable
20 machine a mixture of the powdered compound moistened with an inert
liquid diluent. The tablets may optionally be coated or scored and may be
formulated so as to provide slow or controlled release of the active
ingredient therein using, for example, hydroxypropylmethylcellulose in
varying proportions to provide desired release profile.

25 Formulations suitable for topical administration in the mouth include
lozenges comprising the active ingredient in a flavoured basis, usually
sucrose and acacia or tragacanth; pastilles comprising the active ingredient
in an inert basis such as gelatin and glycerin, or sucrose and acacia; and
30 mouth-washes comprising the active ingredient in a suitable liquid carrier.

In human therapy, the compounds of the invention can be administered alone but will generally be administered in admixture with a suitable pharmaceutical excipient diluent or carrier selected with regard to the intended route of administration and standard pharmaceutical practice.

5

For example, the compounds of the invention can be administered orally, buccally or sublingually in the form of tablets, capsules, ovules, elixirs, solutions or suspensions, which may contain flavouring or colouring agents, for immediate-, delayed- or controlled-release applications. The
10 compounds of the invention may also be administered *via* intracavernosal injection.

Such tablets may contain excipients such as microcrystalline cellulose, lactose, sodium citrate, calcium carbonate, dibasic calcium phosphate and
15 glycine, disintegrants such as starch (preferably corn, potato or tapioca starch), sodium starch glycollate, croscarmellose sodium and certain complex silicates, and granulation binders such as polyvinylpyrrolidone, hydroxypropylmethylcellulose (HPMC), hydroxy-propylcellulose (HPC), sucrose, gelatin and acacia. Additionally, lubricating agents such as
20 magnesium stearate, stearic acid, glyceryl behenate and talc may be included.

Solid compositions of a similar type may also be employed as fillers in gelatin capsules. Preferred excipients in this regard include lactose, starch,
25 cellulose, milk sugar or high molecular weight polyethylene glycols. For aqueous suspensions and/or elixirs, the compounds of the invention may be combined with various sweetening or flavouring agents, colouring matter or dyes, with emulsifying and/or suspending agents and with diluents such as water, ethanol, propylene glycol and glycerin, and combinations thereof.

30

The compounds of the invention can also be administered parenterally, for example, intravenously, intra-arterially, intraperitoneally, intra-theCALLY, intraventricularly, intrasternally, intracranially, intra-muscularly or subcutaneously, or they may be administered by infusion techniques. They
5 are best used in the form of a sterile aqueous solution which may contain other substances, for example, enough salts or glucose to make the solution isotonic with blood. The aqueous solutions should be suitably buffered (preferably to a pH of from 3 to 9), if necessary. The preparation of suitable parenteral formulations under sterile conditions is readily accomplished by
10 standard pharmaceutical techniques well-known to those skilled in the art.

Formulations suitable for parenteral administration include aqueous and non-aqueous sterile injection solutions which may contain anti-oxidants, buffers, bacteriostats and solutes which render the formulation isotonic with the blood
15 of the intended recipient; and aqueous and non-aqueous sterile suspensions which may include suspending agents and thickening agents. The formulations may be presented in unit-dose or multi-dose containers, for example sealed ampoules and vials, and may be stored in a freeze-dried (lyophilised) condition requiring only the addition of the sterile liquid carrier,
20 for example water for injections, immediately prior to use. Extemporaneous injection solutions and suspensions may be prepared from sterile powders, granules and tablets of the kind previously described.

Generally, in humans, oral or parenteral administration of the compounds of
25 the invention is the preferred route, being the most convenient.

For veterinary use, the compounds of the invention are administered as a suitably acceptable formulation in accordance with normal veterinary practice and the veterinary surgeon will determine the dosing regimen and
30 route of administration which will be most appropriate for a particular animal.

The formulations of the pharmaceutical compositions of the invention may conveniently be presented in unit dosage form and may be prepared by any of the methods well known in the art of pharmacy. Such methods include
5 the step of bringing into association the active ingredient with the carrier which constitutes one or more accessory ingredients. In general the formulations are prepared by uniformly and intimately bringing into association the active ingredient with liquid carriers or finely divided solid carriers or both, and then, if necessary, shaping the product.

10

Preferred unit dosage formulations are those containing a daily dose or unit, daily sub-dose or an appropriate fraction thereof, of an active ingredient.

A preferred delivery system of the invention may comprise a hydrogel
15 impregnated with a compound of the invention, which is preferably carried on a tampon which can be inserted into the cervix and withdrawn once an appropriate cervical ripening or other desirable affect on the female reproductive system has been produced.

20 It should be understood that in addition to the ingredients particularly mentioned above the formulations of this invention may include other agents conventional in the art having regard to the type of formulation in question, for example those suitable for oral administration may include flavouring agents.

25

EXAMPLE 3 – Exemplary pharmaceutical formulations

Whilst it is possible for a compound of the invention to be administered alone, it is preferable to present it as a pharmaceutical formulation, together with one
5 or more acceptable carriers. The carrier(s) must be “acceptable” in the sense of being compatible with the compound of the invention and not deleterious to the recipients thereof. Typically, the carriers will be water or saline which will be sterile and pyrogen-free.

10 The following examples illustrate pharmaceutical formulations according to the invention in which the active ingredient is a compound of the invention.

Example 3A: Tablet

15	Active ingredient	100 mg
	Lactose	200 mg
	Starch	50 mg
	Polyvinylpyrrolidone	5 mg
	Magnesium stearate	4 mg
20		359 mg

Tablets are prepared from the foregoing ingredients by wet granulation followed by compression.

25

Example 3B: Ophthalmic Solution

	Active ingredient	0.5 g
	Sodium chloride, analytical grade	0.9 g
5	Thiomersal	0.001 g
	Purified water to	100 ml
	pH adjusted to	7.5

Example 3C: Tablet Formulations

10

The following formulations A and B are prepared by wet granulation of the ingredients with a solution of povidone, followed by addition of magnesium stearate and compression.

15 *Formulation A*

	<u>mg/tablet</u>	<u>mg/tablet</u>
(a) Active ingredient	250	250
(b) Lactose B.P.	210	26
(c) Povidone B.P.	15	9
20 (d) Sodium Starch Glycolate	20	12
(e) Magnesium Stearate	5	3
	—	
	500	300

25 *Formulation B*

	<u>mg/tablet</u>	<u>mg/tablet</u>
(a) Active ingredient	250	250
(b) Lactose	150	-
(c) Avicel PH 101 [®]	60	26
30 (d) Povidone B.P.	15	9
(e) Sodium Starch Glycolate	20	12

(f) Magnesium Stearate	5	3
	—	
	500	300

5 *Formulation C*

	<u>mg/tablet</u>
Active ingredient	100
Lactose	200
Starch	50
10 Povidone	5
Magnesium stearate	4
	359

- 15 The following formulations, D and E, are prepared by direct compression of the admixed ingredients. The lactose used in formulation E is of the direction compression type.

Formulation D

	<u>mg/capsule</u>
20 Active Ingredient	250
Pregelatinised Starch NF15	150
	400

25 *Formulation E*

	<u>mg/capsule</u>
Active Ingredient	250
Lactose	150
Avicel [®]	100
30	500

Formulation F (Controlled Release Formulation)

The formulation is prepared by wet granulation of the ingredients (below)
 5 with a solution of povidone followed by the addition of magnesium stearate
 and compression.

	<u>mg/tablet</u>
Active Ingredient	500
Hydroxypropylmethylcellulose	112
10 (Methocel K4M Premium) [®]	
Lactose B.P.	53
Povidone B.P.C.	28
Magnesium Stearate	7
15	700

Drug release takes place over a period of about 6-8 hours and was complete
 after 12 hours.

Example 3D: Capsule Formulations

20

Formulation A

A capsule formulation is prepared by admixing the ingredients of
 Formulation D in Example C above and filling into a two-part hard gelatin
 25 capsule. Formulation B (*infra*) is prepared in a similar manner.

Formulation B

	<u>mg/capsule</u>
Active ingredient	250
30 Lactose B.P.	143
Sodium Starch Glycolate	25

Magnesium Stearate 2

420

5 *Formulation C*

	<u>mg/capsule</u>
Active ingredient	250
Macrogol 4000 BP	350

10 600

Capsules are prepared by melting the Macrogol 4000 BP, dispersing the active ingredient in the melt and filling the melt into a two-part hard gelatin capsule.

15 *Formulation D*

	<u>mg/capsule</u>
Active ingredient	250
Lecithin	100
Arachis Oil	100

20

450

Capsules are prepared by dispersing the active ingredient in the lecithin and arachis oil and filling the dispersion into soft, elastic gelatin capsules.

25

Formulation E (Controlled Release Capsule)

The following controlled release capsule formulation is prepared by extruding ingredients a, b, and c using an extruder, followed by spheronisation of the extrudate and drying. The dried pellets are then coated with release-controlling membrane (d) and filled into a two-piece, hard gelatin capsule.

30

	<u>mg/capsule</u>
Active ingredient	250
Microcrystalline Cellulose	125
Lactose BP	125
5 Ethyl Cellulose	13
	513

Example 3E: Injectable Formulation

10

Active ingredient 0.200 g
Sterile, pyrogen free phosphate buffer (pH7.0) to 10 ml

15 The active ingredient is dissolved in most of the phosphate buffer (35-40°C), then made up to volume and filtered through a sterile micropore filter into a sterile 10 ml amber glass vial (type 1) and sealed with sterile closures and overseals.

Example 3F: Intramuscular injection

20

Active ingredient 0.20 g
Benzyl Alcohol 0.10 g
Glucofurol 75[®] 1.45 g
Water for Injection q.s. to 3.00 ml

25

The active ingredient is dissolved in the glycofurol. The benzyl alcohol is then added and dissolved, and water added to 3 ml. The mixture is then filtered through a sterile micropore filter and sealed in sterile 3 ml glass vials (type 1).

30

Example 3G: Syrup Suspension

	Active ingredient	0.2500 g
	Sorbitol Solution	1.5000 g
5	Glycerol	2.0000 g
	Dispersible Cellulose	0.0750 g
	Sodium Benzoate	0.0050 g
	Flavour, Peach 17.42.3169	0.0125 ml
	Purified Water q.s. to	5.0000 ml

10

The sodium benzoate is dissolved in a portion of the purified water and the sorbitol solution added. The active ingredient is added and dispersed. In the glycerol is dispersed the thickener (dispersible cellulose). The two dispersions are mixed and made up to the required volume with the purified water.

15 Further thickening is achieved as required by extra shearing of the suspension.

Example 3H: Suppository

		<u>mg/suppository</u>
	Active ingredient (63 μ m)*	250
20	Hard Fat, BP (Witepsol H15 - Dynamit Nobel)	1770
		2020

25 *The active ingredient is used as a powder wherein at least 90% of the particles are of 63 μ m diameter or less.

One fifth of the Witepsol H15 is melted in a steam-jacketed pan at 45°C maximum. The active ingredient is sifted through a 200 μ m sieve and added to the molten base with mixing, using a silverson fitted with a cutting head,
 30 until a smooth dispersion is achieved. Maintaining the mixture at 45°C, the remaining Witepsol H15 is added to the suspension and stirred to ensure a

homogenous mix. The entire suspension is passed through a 250 µm stainless steel screen and, with continuous stirring, is allowed to cool to 40°C. At a temperature of 38°C to 40°C 2.02 g of the mixture is filled into suitable plastic moulds. The suppositories are allowed to cool to room temperature.

5

Example 3I: Pessaries

	<u>mg/pessary</u>
Active ingredient	250
Anhydrate Dextrose	380
10 Potato Starch	363
Magnesium Stearate	7
	1000

- 15 The above ingredients are mixed directly and pessaries prepared by direct compression of the resulting mixture.

Example 3J: Creams and ointments

- 20 Described in *Remington*.

Example 3K: Microsphere formulations

- 25 The compounds of the invention may also be delivered using microsphere formulations, such as those described in Cleland (1997; 2001).

Example 3L: Dry Powder Inhalation

- 30 The compounds of the invention may be delivered by inhalation, with the aid of a dry powder inhaler delivering micronised particles in metered quantities as described in Ansel (1999).

Example 3M: Aerosol Inhalation

5 The compounds of the invention may be delivered by inhalation, with the aid of a suitable inhaler delivering micronised particles in metered quantities employing a non CFC propellant as described in Ansel (1999).

References

- Allport VC, Pieber D, Slater DM, Newton R, White JO, Bennett PR. Human labour is associated with nuclear factor- κ B activity which mediates cyclo-oxygenase-2 expression and is involved with the functional progesterone withdrawal, *Mol Hum Reprod* 2001; 7: 581-586.
- Ansel. Pharmaceutical Dosage Forms and Drug Delivery Systems, 1999, Lippincott Williams and Wilkins.
- Bendixen AC, Shevde NK, Dienger KM, Willson TM, Funk CD, Pike JW. IL-4 inhibits osteoclast formation through a direct action on osteoclast precursors via peroxisome proliferator-activated receptor γ 1. *Proc Natl Acad Sci USA* 2001; 98: 2443-2448.
- Bennett PR, Rose MP, Myatt L, Elder MG. Preterm labor: stimulation of arachidonic acid metabolism in human amnion cells by bacterial products. *Am J Obstet Gynecol*. 1987, 156:649-5.
- Bennett, P. and Slater, D. The role of cyclo-oxygenases in the onset of labour., in improved non-steroid anti-inflammatory drugs: COX-2 enzyme inhibitors. J. Vane, R. Botting, and G. Botting, Editors. 1996, Kluwer Academic: London. P. 112-118.
- Brown, N.L., Alvi, S.A., Elder, M.G., Bennett, P.R. and Sullivan, M.H. A spontaneous induction of fetal membrane prostaglandin production precedes clinical labour. *J Endocrinol* 1998; 157: R1-R6.
- Carlson *et al.*, *Obstet Gynecol*, 1992. 85(5), 769-774.

Chinetti G, Griglio S, Antonucci M, Torra IP, Delerive P, Majd Z, Fruchart J-C, Chapman J, Najib J, Staels B. Activation of proliferator-activated receptors α and γ induces apoptosis of human monocyte-derived macrophages. *J Biol Chem* 1998; 273: 25573-25580.

5

Cleland, J.L. (1997) *Pharm. Biotechnol.* 10:1-43.

Cleland *et al.* (2001) *J. Control. Release* 72:13-24.

- 10 Corey, E. J. The logic of chemical synthesis-multistep synthesis of complex carbogenic molecules. *Angew Chem Int Ed Engl*, 1991, 30:455-465.

- Crankshaw, D.J. and Dyal, R. Effects of some naturally occurring prostanoids and some cyclo-oxygenase inhibitors on the contractility of the
15 human lower uterine segment *in vitro*. *Can J Physiol Pharmacol*, 1994, 72(8): p. 870-4.

- Dammann O and Leviton A. Brain damage in preterm newborns: Might enhancement of developmentally regulated endogenous protection open a
20 door for prevention? *Pediatrics* 1999; 104: 541-550.

Derossi *et al.* (1998), *Trends Cell Biol* 8, 84-87.

- Dignam *et al.*, Accurate transcription initiation by RNA polymerase II in a
25 soluble extract from isolated mammalian nuclei. *Nucleic Acids Res.*, 11:1475-1489.

- Dyal R and Crankshaw DJ. The effects of some synthetic prostanoids on the contractility of the human lower uterine segment *in vitro*. *Am J Obstet
30 Gynecol* 1988; 158: 281-285.

Elliot CL, Loudon JA, Brown N, Slater DM, Bennett PR, Sullivan MH. IL-1beta and IL-8 in human fetal membranes: changes with gestational age, labor, and culture conditions. *Am J Reprod Immunol* 2001; 46: 260-267.

- 5 Ferry G, Bruneau V, Beauverger P, *et al.* 2001 Binding of prostaglandins to human PPARgamma: tool assessment and new natural ligands. *Eur J Pharmacol.*, 417:77-89.

- Forman BM, Tontonoz P, Chen J, Brun RP, Spiegelman BM, Evans RM.
10 15-deoxy- $\Delta^{12,14}$ -prostaglandin J₂ is a ligand for the adipocyte determination factor PPAR γ . *Cell* 1995; 83: 803-812.

Ganstrom *et al.*, *Acta Obstet Gynecol Scand*, 1987. 66:429-431.

- 15 Garfield RE and Hertzberg EL. Cell-to-cell coupling in the myometrium: Emil Bozler's prediction. *Prog Clin Biol Res* 1990; 327: 673-681.

- Gibson CS, MacLennan AH, Goldwater PN, Dekker GA. Antenatal causes of cerebral palsy: associations between inherited thrombophilias, viral and
20 bacterial infection, and inherited susceptibility to infection. *Obstet Gynecol Survey* 2003; 58: 209-220.

- Gupta RA, Polk DB, Krishna U, Israel DA, Yan F, DuBois RN, Peek RM Jr. Activation of peroxisome proliferator-activated receptor γ suppresses
25 nuclear factor κ B-mediated apoptosis induced by *Helicobacter Pylori* I gastric epithelial cells. *J Biol Chem* 2001; 276: 31059-31066.

- Herman A, Groutzd A, Bukovsky I, Arieli S, Sherman D, Caspi E. A simplified pre-induction scoring method for the prediction of successful
30 vaginal delivery based on multivariate analysis of pelvic and other obstetrical factors. *J Perinat Med.* 1993, 21:117-24.

Huang JT, Welch JS, Ricote M, Binder CJ, Willson TM, Kelly C, Witztum JL, Funk CD, Conrad D, Glass CK. Interleukin-4-dependent production of PPAR- γ ligands in macrophages by 12/15-lipoxygenase. *Nature* 1999; 400:
5 378-382.

Ikawa H, Kameda H, Kamitani H, Baek SJ, Nixon JB, His LC, Eling TE. Effect of PPAR activators on cytokine-stimulated cyclooxygenase-2 expression in human colorectal carcinoma cells. *Exp Cell Res* 2001; 267:
10 73-80.

Johnson C, Van Antwerp D, Hope TJ. An N-terminal nuclear export signal is required for the nucleocytoplasmic shuttling of I κ B α . *EMBO J* 1999; 23:
15 6682-6693.

Keelan JA, Marvin KW, Sato TA, Coleman M, McCowan LM, Mitchell MD. Cytokine abundance in placental tissues: evidence of inflammatory activation in gestational membranes with term and preterm parturition. *Am J Obstet Gynecol* 1999; 181: 1530-1536.
20

Keirse (1995) "Indomethacin tocolysis in pre-term labour" in *Pregnancy and Childbirth Module* (Eds. Enkin, M.W., Keirse, M.J.N.C., Renfrew, M.J., Neilson, J.P.) Cochrane Database of Systematic Reviews, No 04383, Oxford).

25 Kelly RW. Inflammatory mediators and cervical ripening. *J Reprod Immunol* 2002; 57: 217-224.

Kliwer SA, Lenhard JM, Willson TM, Patel I, Morris DC, Lehmann JM
30 1995 A prostaglandin J2 metabolite binds peroxisome proliferator-activated receptor gamma and promotes adipocyte differentiation. *Cell* 83:813-9.

Kunsch C, Ruben SM, Rosen CA. Selection of optimal kappa B/Rel DNA-binding motifs: interaction of both subunits of NF-kappaB with DNA is required for transcriptional activation. *Mol Cell Biol* 1992; 12: 4412-4421.

5

Lee, *Curr. Opin. Biotechnol.*, 2001, 11:81-84.

Lee Y, Allport V, Sykes A, Lindstrom T, Slater D, Bennett P. The effects of labour and of interleukin 1 beta upon the expression of nuclear factor
10 kappa B related proteins in human amnion. *Mol Hum Reprod* 2003; 9: 213-8.

Leesnitzer LM, Parks DJ, Bledsoe RK, Cobb JE, Collins JL, Consler TG, Davis RG, Hull-Ryde EA, Lenhard JM, Patel L, Plunket KD, Shenk JL,
15 Stimmel JB, Therapontos C, Willson TM, Blanchard SG. Functional consequences of cysteine modification in the ligand binding sites of peroxisome proliferator activated receptors by GW9662. *Biochem* 2002; 41: 6640-50.

20 Li Q and Verma IM. NF- κ B regulation in the immune system. *Nature Reviews*, 2002. 2:725-735.

Lin R, Gewert D, Hiscott J. Differential transcriptional activation in vitro by NF- κ B/Rel proteins. *J Biol Chem* 1995; 270: 3123-3131.

25

Maul H, Nagel S, Welsch G, Schafer A, Winkler M, Rath W. Messenger ribonucleic acid levels of interleukin-1 beta, interleukin-6 and interleukin-8 in the lower uterine segment increased significantly at final cervical dilatation during term parturition, while those of tumor necrosis factor alpha
30 remained unchanged. *Eur J Obstet Gynecol Reprod Biol* 2002; 102:143-7.

Meade EA, McIntyre TM, Zimmerman GA, Prescott SM. Peroxisome proliferators enhance cyclooxygenase-2 expression in epithelial cells. *J Biol Chem* 1999; 274: 8328-8334.

5 Mitchell MD, Edwin SS, Lundin-Schiller S, Silver RM, Smotkin D, Trautman MS. Mechanism of interleukin-1 beta stimulation of human amnion prostaglandin biosynthesis: mediation via a novel inducible cyclooxygenase. *Placenta* 1993; 14: 615-625.

10 Miyahara T, Schrum L, Rippe R, Xiong S, Yee HF Jr, Motomura K, Anania FA, Willson TM, Tsukamoto H. Peroxisome proliferator-activated receptors and hepatic stellate cell activation. *J Biol Chem* 2000; 275: 35715-35722.

15 Moise *et al.* Effect of advancing gestational age on the frequency of fetal ductal constriction in association with maternal indomethacin use" *Am. J. Obstet. Gynecol.*, 1995, 170(45), 1204-5.

Narumiya S. Structures, properties and distributions of prostanoid
20 receptors. *Adv Prost Thromb Leuk Res* 1995; 23: 17-22.

Narumiya S, Ohno K, Fukushima M, Fujiwara M. Site and mechanism of growth inhibition by prostaglandins. III. Distribution and binding of prostaglandin A2 and delta 12-prostaglandin J2 in nuclei. *J Pharmacol Exp*
25 *Ther* 1987; 242: 306-11.

Nasuhura *et al.*, *JBC*, 1999. 274:19965.

Pang L, Nie M, Corbett L, Knox AJ. Cyclooxygenase-2 expression by
30 nonsteroidal anti-inflammatory drugs in human airway smooth muscle cells:

Role of peroxisome proliferator-activated receptors. *J Immunol* 2003; 170: 1043-1051.

Phelps CB, Sengchanthalangsy LL, Malek S, Ghosh G. Mechanism of κ B
5 DNA binding by Rel/NF- κ B dimers. *J Biol Chem* 2000; 275: 24392-24399.

Pieber D, Allport VC, Hills F, Johnson M, Bennett PR. Interactions between
progesterone receptor isoforms in myometrial cells in human labour. *Mol*
Hum Reprod. 2001, 7:875-9.

10 Prusty D, Park B-H, Davis KE, Farmer SR. Activation of MEK/ERK
signaling promotes adipogenesis by enhancing peroxisome proliferators-
activated receptor γ (PPAR γ) and C/EBP α gene expression during the
differentiation of 3T3-L1 preadipocytes. *J Biol Chem* 2002; 277: 46226-
15 46232.

Rasanen and Jouppila. Fetal cardiac function and ductus arteriosus during
indomethacin and sulindac therapy for threatened pre-term labour; A
randomised study. *Am J Obstet Gynecol* 1995. 173(1), 20-25.

20 *Remington: The Science and Practise of Pharmacy*, 19th ed., The
Philadelphia College of Pharmacy and Science, ISBN 0-912734-04-3.

Respondek *et al.*, Fetal echocardiography during indomethacin treatment.
25 *Ultrasound Obstet Gynecol*, 1995. 5, 86-89.

Romero R, Parvizi ST, Oyarzun E, Mazor M, Wu YK, Avila C,
Athanassiadis AP, Mitchell MD. Amniotic fluid interleukin-1 in
spontaneous labour at term. *J Reprod Med* 1990; 35: 235-238.

30

- Romero R, Espinoza J, Chaiworapongsa T, Kalache K. Infection and prematurity and the role of preventive strategies. *Semin Neonatol.* 2002; 7:259-74.
- 5 Ruan H, Pownall HJ, Lodish HF. Troglitazone antagonizes TNF- α -induced reprogramming of adipocyte gene expression by inhibiting the transcriptional regulatory functions of NF- κ B. *J Biol Chem* 2003; Manuscript M303141200.
- 10 Rush RW, Keirse MJNC, Howat P, Baum JD, Anderson AB, Turnbull AC. Contribution of preterm delivery to perinatal mortality. *Br Med J* 1976; 2: 965.
- Satoh K, Yasumizu T, Fukuoka H, Kinoshita K, Kaneko Y, Tsuchiya M, 15 Sakamoto S. Prostaglandin F2 alpha metabolite levels in plasma, amniotic fluid, and urine during pregnancy and labor. *Am J Obstet Gynecol* 1979; 133: 886-890.
- Sambrook *et al.*, Molecular Cloning. A laboratory manual. 1989. Cold 20 Spring Harbour pub.
- Schreiber *et al.*, Rapid detection of octamer binding proteins with mini-extracts prepared from a small number of cells. *Nucl. Acids Res*, 1989. 17:6419.
- 25 Skinner KA and Challis JR. Changes in the synthesis and metabolism of prostaglandins by human fetal membranes and decidua at labour. *Am J Obstet Gynecol* 1985; 151: 519-523.

- Slater DM, Berger L, Newton R, Moore GE, Bennett PR. Changes in the expression of types 1 and 2 cyclo-oxygenase in human fetal membranes at term. *Am J Obstet Gynecol* 1995; 172: 77-82.
- 5 Staels B, Koenig W, Habib A, Merval R, Lebreton M, Torra IP, Delerive P, Fadel A, Chinetti G, Fruchart JC, Najib J, Macclouf J, Tedgui A. Activation of human aortic smooth-muscle cells is inhibited by PPAR α but not by PPAR γ activators. *Nature* 1998; 393: 790-793.
- 10 Straus D. S. and Glass C. K. Cyclopentenone prostaglandins: new insights on biological activities and cellular targets. *Med Res Rev*, 2001. 21:185-210.
- 15 Subbaramaiah K, Lin DT, Hart JC, Dannenberg AJ. Peroxisome proliferator-activated receptor γ ligands suppress the transcriptional activation of cyclooxygenase-2. *J Biol Chem* 2001; 276: 12440-12448.
- 20 Suyang H, Phillips R, Douglas I, Ghosh S. Role of unphosphorylated, newly synthesized I κ B β in persistent activation of NF- κ B. *Mol Cell Biol* 1996; 16: 5444-5449.
- 25 Suzawa M, Takada I, Yanagisawa J, Ohtake F, Ogawa S, Yamauchi T, Kadowaki T, Takeuchi Y, Shibuya H, Gotoh Y, Matsumoto K, Kato S. Cytokines suppress adipogenesis and PPAR- γ function through the TAK1/TAB1/NIK cascade. *Nature Cell Biol* 2003; 5: 224-230.
- Suzuki *et al.*, Three component coupling synthesis of prostaglandins. A simplified, general procedure. *Tetrahedron*, 1990, 46:4809-4822.
- 30 Takata Y, Kitami Y, Yang Z-H, Nakamura M, Okura T, Hiwada K. Vascular inflammation is negatively autoregulated by interaction between

CCAAT/enhancer-binding protein- δ and peroxisome proliferator-activated receptor- γ . *Circ Res* 2002; 91: 427-433.

Takeuchi *et al.* *Adv. Drug. Delic. Rev.*, 2001, 47:39-54.

5

Tanaka T, Itoh H, Doi K, Fukunaga Y, Hosoda K, Shintani M, Yamashita J, Chun TH, Inoue M, Masatsugu K, Sawada N, Saito T, Inoue G, Nishimura H, Yoshimasa Y, Nakao K. Down regulation of peroxisome proliferator-activated receptor γ expression by inflammatory cytokines and its reversal by thiazolidinediones. *Diabetologia* 1999; 42: 702-710.

10

Toledano MB, Ghosh D, Trinh F, Leonard WJ. N-terminal DNA-binding domains contribute to differential DNA-binding specificities of NF-kappa B p50 and p65. *Mol Cell Biol* 1993; 13: 852-860.

15

Tontonoz P, Nagy L, Alvarez J, Thomazy V, Evans R. PPAR γ promotes monocyte/macrophage differentiation and uptake of oxidized LDL. *Cell* 1998; 93: 241-252.

20 Tulzer *et al.*, Doppler- echocardiography of fetal ductus arteriosus constriction versus increased right ventricular output. *JACC*, 1991. 18(2), 532-36.

Turnbull, A. The fetus and birth, in Elsevier, London. 1977.

25

Urakubo A, Jarskog LF, Lieberman JA, Gilmore JH. Prenatal exposure to maternal infection alters cytokine expression in the placenta, amniotic fluid, and fetal brain. *Schizophrenia research* 2001; 47: 27-36.

Van Meir, C.A., *et al.*, Chorionic prostaglandin catabolism is decreased in the lower uterine segment with term labour. *Placenta*, 1997. 18(2-3): p.109-14.

- 5 Van Meir, C.A., *et al.*, Immunoreactive 15-hydroxyprostaglandin dehydrogenase (PGDH) is reduced in fetal membranes from patients at pre-term delivery in the presence of infection. *Placenta*, 1996. 17(5-6): p.291-7.

Volpe JJ. Neurobiology of periventricular leukomalacia in the premature
10 infant. *Pediatr Res* 2001;50:553-62.

Ward C, Dransfield I, Murray J, Farrow SN, Haslett C, Rossi AG. Prostaglandin D₂ and its metabolites induce caspase-dependent granulocyte apoptosis that is mediated via inhibition of I κ B α degradation using a
15 peroxisome proliferator-activated receptor- γ -independent mechanism. *J Immunol* 2002; 168: 6232-6243.

Whiteside ST, Epinat J-C, Rice NR, Israel A. I kappa B epsilon, a novel member of the I κ B family, controls RelA and cRel NF- κ B activity. *EMBO J* 1997(b); 16: 1413-1426.
20

Zhou YC and Waxman DJ. Cross-talk between Janus-Kinase-signal transducer activator of transcription (JAK-STAT) and peroxisome proliferator-activated α (PPAR α) signaling pathways. *J Biol Chem* 1999;
25 274: 2672-2681.